# UNIVERSITY OF BENGHAZI FACULTY OF SCIENCE CHEMISTRY DEPARTMENT



# This Thesis Entitled

# THE PYRROLIZIDINE ALKALOIDS OF

# ECHIUM PLANTAGENIUM L

By

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Submitted in Partial Fulfillment for Requirement for Master Science Degree in Chemistry

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July (2015)

جامعة بنغازي كاية العلوم قسم الكيمياء



أطروحة ماجستير بعنوان

# قلويدات البروليزيدين في نبات الأيشيم بلانتاجنيم (Echium Plantagenium L)

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# $\lq\lq$ The Pyrrolizidine alkaloids of *Echium Plantagenium* $L\lq\lq$

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#### LIST OF ABBREVIATIONS

Ac Acetyl
Ang Anoeloyl
Bu Butyl

CRO Crotanecine
Ech Echimidinoyl

Ech-number Alkaloid-Compound Number

EI Electron Impact

EM Electron Multiplier

EtOH Ethanol

GC Gas Chromatography

GC-MS Gas Chromatography Mass Spectrometry

GLC Gas Liquid Chromatography

HEL Heliotridine

HPLC High performance Liquid Chromatography

Int Interrinecoyl
Las Lasiocapoyl
MeOH Methanol

MS Mass Spectrometry

NMR Nuclear Magnetic Resonance

OTO otonecine

PAs Pyrrolizidine alkaloids

Ph Phenyl

RET Retronecine

Rf Retention Factor
RI Retention Indice

Rt Retention time

Sar Sarrascinoyl

Sen Senecioyl
SUP Supinidine
Tig Tigloyl

TLC Then Layer Chromatography

Tra Trachelanthoyl

Vir viridifloyl

#### **ABSTRACT**

This study includes brief review of previously isolated and identified pyrrolizidine alkaloids from various Boraginaceae plant species by using various isolation and characterization techniques.

The primary aim of this research was isolation and characterization of pyrrolizidine alkaloids from *Echium plantagenium* (Boraginaceae); an uninvestigated Libyan species which grown around El marj city.

Gas Chromatographic Mass Spectrometry technique were used for isolation of pyrrolizidine alkaloids from Methanolic extraction of *Echium plantagenium*. Which result in isolate a twelve compounds. The chemical structures of the isolated compounds have been elucidated by using their Mass spectrometric fragmentation pattern and the value of retention indices; in compare with those of previously identified pyrrolizidine alkaloids as reference standard values.

A number of isolated compounds were tentatively identified; while the rest of them is not elucidated accurately, but has been proposed a number of isomers to represent their chemical structure as follow:

Lycopsamine

Assumed one of the following isomers

$\mathbf{R}_1$	$R_2$	$R_3$	
ОН	OAc	Н	3'-acetylintermedine
OAc	ОН	Н	2'-acetylintermedine
ОН	Н	OAc	3'-acetylycopsamine
OAc	Н	ОН	2'-acetylycopsamine

Ech-7

7-Angeloyl-9-(2,3-dihydroxybutyryl) retronecine

Ech-8 and Ech-9	Ech-10
Assumed to be isomer of Ech-7	O HO O HO
	Uplandicine
Ech-11	Ech-12
HO HO OH HO Echimidine	HO OH O

#### **ACKNOWLEDGEMENT**

I offer the unlimited thanks to ALLAH in making me have the momentum to meet all efforts required during the course of this work.

I would like to express my deep feeling of gratitude, great indebtedness and sincerer appreciation to Prof. Dr. Fakhri A.Elabbar, Chemistry Department, Faculty of science, University of Benghazi for his kind supervision, helpful guidance and continuous encouragement during the course of this work.

I wish also to express my grateful acknowledgement to Ms Abdo Slam El-Mogasapi Department of Botany, Faculty of Science, University of Benghazi for valuable help in the collection and identification of plant material.

I wish to express my sincere thanks to my colleagues in University of Benghazi and for their help, continuous encouragement and understanding.

Finally, my cordial appreciation and deepest thanks to my family specially my wife for their patience, sincere help, understanding and continuous encouragement.

The Auther

#### **CHAPTER ONE**

#### 1. Introduction

#### 1.1. Natural Products

By definition, natural products are compounds present in or produced by nature and not artificial or man-made[1].

Naturally occurring compounds may be divided into three broad categories. Firstly, primary metabolites compounds which occur in all cells and play a central role in the metabolism and reproduction of those cells. such as the nucleic acids and the common amino acids and sugars. Secondly, high-molecular-weight polymeric materials such as cellulose, the lignins and the proteins which form the cellular structures. Thirdly, there are the secondary metabolites compounds that are not essential to the growth and development of the producing organism, and characteristic of a limited range of species[2].

The term natural product really refers to any naturally occurring substance but is generally taken to mean a secondary metabolites which Cannot be divided in a clear manner because of the large variation in their structures[3].

Scientists working with natural products with interest for the purpose of and elucidation of their structures and their chemistry or characterization of their biological effects

#### 1.2. Alkaloids

Alkaloids are secondary metabolites compounds with low molecular weight present mainly in many families of the plant kingdom, and they are produced by a large variety of organisms including microbiology organism (e.g. bacteria, fungi), plants, and animals[4-6].

The term alkaloid, coined in 1819 in Halle, Germany, by the pharmacist, Carl Meissner, finds its origin in the Arabic name *al-qali*, the plant from which soda was first isolated[7].

Ladenburg defined alkaloids as naturally occurring plant compounds having a basic character and containing at least one nitrogen in a heterocyclic ring.' With the advent of recent advanced knowledge in the chemistry of various alkaloids two more characteristic features were logically and justifiably added to the definition of alkaloids, namely:(a) Complex

molecular structure, (b) Significant pharmacological activity. Furthermore, it was broadly observed that the basic properties of the alkaloids is solely by virtue of the presence of N-atom embedded into the ring system[8].

Therefore, the alkaloids are now generally defined as, physiologically active basic compounds of plant origin, in which at least one nitrogen atom forms part of a cyclic system.' Even this definition has a few anomalies as stated below[8]:

(i) Cholines and Betaines: These two substances have the N-atom in the side chain and not in the aromatic ring as shown below:

HO 
$$\uparrow$$
  $N(CH_3)_3$   $(H_3C)_3N$   $O$ 

Choline Betaine

The cholines and betaines are regarded as simple alkylamines and not classified as alkaloids. They are designated by some school of thoughts as 'biological-amines' or 'protoalkaloids'.

(ii) Ephedrine: It has the N-atom only in the side chain and not embedded in the aromatic ring as given below:

**Ephedrine** 

(iii) Piperidine: It is obtained Piper nigrum (Black Pepper) and does not possess any pharmacological activity, but has a N-atom in a heterocyclic ring as given below:

#### Piperidine

(iv) Colchicine: It is found to be neither basic nor it contains the N-atom in a heterocyclic ring, whereas it is considered as an alkaloid due to the fact it possesses distinct pharmacological activity as shown below:

Cholchcine

(v) Thiamine (Vitamin B1): It confines to the definition of alkaloids but is not regarded as an'alkaloid' because of its almost universal distribution in living matter.

Thiamine monochloride

More than 27000 structure of alkaloid has been diagnosed from variety plant species. A single plant species usually contains only a few kinds of alkaloids, but certain plant families may be rich in alkaloids[9].

Alkaloids They includes in their structure one nitrogen atom or more typically in the form of a primary, secondary, or tertiary amines; that's what makes them bases[10, 11]. Many alkaloids which are toxic to other organisms can be purified from crude extracts by acid - base extraction[12].

Many alkaloids have a wide range of pharmacological activities and found use in traditional or modern medicine, or as starting points for drug discovery. Other alkaloids possess psychotropic (e.g. psilocin) and stimulant activities (e.g. cocaine)[13], Alkaloids can be toxic too (e.g. atropine, tubocurarine). Although alkaloids act on a diversity of metabolic systems in humans and other animals, they almost uniformly evoke a bitter taste[8].

#### 1.2.1. General Characteristics of Alkaloids

#### 1.2.1.1. The basicity of alkaloids

The alkaloids are organic nitrogenous bases found mainly in plants. They have one or more nitrogen atoms present, typically as primary, secondary, or tertiary amines, and this usually confers basicity to the alkaloid, facilitating their isolation and purification since water-soluble salts can be formed in the presence of mineral acids[11].

the degree of basicity varies greatly, depending on the structure of the alkaloid molecule, and the presence and location of other functional groups.45 However, most alkaloids have basic properties with pH value between 6-7[11, 14]. and present in plants in the form of free bases, combined with acids to be salts, or in form of N-oxides. free alkaloids are soluble in organic solvents but mingy water solubility. Alkaloidal salts are soluble in water but insoluble in organic solvents. This difference in solubility provides an excellent way to separate the alkaloids from their natural sources[15].

The alkaloids are usually neutrallized with acids to form salts that may be converted to the corresponding free-base by the cautious addition of selective weak bases, such as, ammonia, calcium hydroxide or sodium carbonate. The usage of either NaOH or KOH solutions must be avoided to prevent the decomposition or destruction of highly sensitive alkaloids[16].

#### 1.2.1.2. The solubility alkaloids

The solubility of different alkaloids and their respective salts usually exhibit considerable variation, which may be attributed from their extremely complex and varied chemical structures. However, it has been observed that the free alkaloid bases as such are invariably found to be fairly soluble in organic solvents, such as: ether, chloroform, relatively non-polar solvents (hexane, benzene, petroleum ether), immiscible solvent, lower alcohols (methanol, ethanol); but they are either practically insoluble soluble in water. Whereas the alkaloidal salts are almost freely soluble in water, relatively less soluble in alcohol and mostly either insoluble or sparingly soluble in organic solvents[8].

#### 1.2.2. Stability of alkaloids

Alkaloid, are not very stable. They normally undergo degradation or decomposition on being exposed to air, light, moisture and heat, besides chemical reagents. For example an aqueous solution of alkaloids undergo rapid decomposition or degradation as compared to their solid forms[16].

During the course of extraction of alkaloids followed by isolation, the solvent is preferably removed effectively by distillation under vacuum (or reduced atmospheric pressure) or by subjecting it to evaporation in a Rotary Thin-Film Evaporator under vacuum so that the desired product is not exposed to excessive heat[17], thus avoiding decomposition.

#### 1.2.3. Testing and precipitating of alkaloids

A good number of alkaloids obtained from various plant sources invariably give a distinct precipitate with certain specific reagents to an extent as small as one microgram. Based on these observations, these alkaloid-precipitating reagents are sometimes employed for either detecting the presence or absence of alkaloids in a crude extracts or plant materials, and for ascertaining whether a specific extraction procedure has exhausted completely the alkaloidal contents or not[8].

Most alkaloids are precipitated from neutral or slightly acid solution by Mayer's reagent (potassio mercuric iodide solution), by Wagner's reagent (solution of iodine in potassium iodide), by solution of tannic acid, by Hager's reagent (a saturated solution of picric acid), or by Dragendorff's reagent (solution of potassium bismuth iodide). These precipitates may be amorphous or crystalline and are of various colours: cream (Mayer's), yellow (Hager's), reddish-brown (Wagner's and Dragendorff's)[15].

The table (1) different qualitative chemical tests can be performed for establishing profile of given extract for its chemical composition.

Name of reagent	Composition
Mayer's Reagent[18]	Mercuric chloride = 1.36 g
(Potassium-Mercuric Iodide Test	Potassium Iodide = 5.00 g
Solution)	Distilled water to make = 100.00 ml

Wagner's Reagent[18]	Iodine = 1.27 g
(Potassium Triiodide)	Potassium = 2.0 g
	Distilled water to make = 100.00 ml
Dragendorff Reagent[14]	Solution A
	55.5 mL of 36% HCl
	bismuth nitrate = 2 g
	Distilled water to make = 100.00 ml
	Solution B
	potassium iodide = 32 g
	Dragendorff reagent prepared by the addition of 20
	mL of solution (A) and 20 mL of solution (B) then
	complete to 100 ml with distilled water
Kraut's Reagent [8]	Bismuth Nitrate = 8.0 g
(Modified Dragendorff'sReagent)	Nitric Acid = 20.0 ml
	Potassium Iodide = 27.2 g
	Distilled water to make = 100.00 ml
Marme's Reagent[8]	Cadmium Iodide = 10.0 g
(Potassium-Cadmium Iodide	Potassium Iodide = 20.0 g
Reagent)	Distilled water to make = 100.00 ml
Scheibler's Reagent [8]	Sodium Tungstate = 20.0 g
(Phosphotungstic Acid Reagent)	Disodium Phosphate = 70.0 g
	Distilled water to make = 100.00 ml
	Note: Acidify with nitric acid to litmus paper.
Hager's reagent[18]	a saturated solution of picric acid
Sonnenschein's Reagent[8]	A 1% (w/v) solution of phosphomolybdic acid in
(Phosphomolybdic Acid	ethanol.
Bertrand's Reagent[8]	A 1% (w/v) solution of silicotungstic acid in
(Silicotungstic Acid)	distilled water.

Ammonium Reineckate = 1.0 g
NH <sub>4</sub> [Cr(NH <sub>3</sub> ) <sub>2</sub> (SCN) <sub>4</sub> ]
Hydroxylamine HCl = 0.3 g
Ethanol = 100.00 ml
Note: Filter and store in a refrigerator.

Table 1: Composition of common reagents used for detection of alkaloids

#### 1.2.4. Classification of Alkaloids

The classification of the alkaloids is complex and may be guided by a set of rules that take into account the structure and other chemical features of the alkaloid molecule, its biological origin, as well as the biogenetic origin where known[19]. some of these rules are describe briefly as below[20]:

**Pharmacological classification** based on the clinical use or pharmacological activity (e.g Cardioactive alkaloids)

**Taxonomic classification** based on family or genus. Without reference to the chemical type of alkaloid present (e.g Solanaceous alkaloids)

**Biosynthetic classification** based on precursors or building block compounds used by plants to synthesize alkaloids (e.g Morphine, papaverine, and colchicines may be listed as phenylanaline and tyrosine derived bases)

**Chemical classification** based on Chemical structure of basic ring in the alkaloid(e.g Atropine is a tropane alkaloid; quinine is a quinoline alkaloid; papaverine is an isoquinoline; strychnine and ergometrine are indol alkaloids)

According to the fact that all alkaloids must contain nitrogen atom at least, alkaloids may be classified based on the nature of the nitrogen-containing structure as follow[21]:

**True alkaloids** that contain nitrogen atom in heterocyclic ring, and derived from amino acid. **Proto alkaloids** or amino alkaloids that does not have nitrogen heterocyclic ring, and derived from amino acid.

**Pseudo alkaloids** contain nitrogen atom in heterocyclic ring, but not derived from amino acid.

Based on chemical nature alkaloids are further classified into two groups broad divisions[15]:

- I . Non-heterocyclic or atypical alkaloids, called 'protoalkaloids' or biological amines.
  - 1. Hordenine or N -methyltyramine
  - 2. Mescaline, related to tryptamine
  - 3. Ephedrine
  - 4. Colchicine (tropolone nucleus with nitrogen in side-chain)
  - 5. Erythromycin (an antibiotic)
  - 6. Jurubin (steroid with 3-amino group)
  - 7. Pachysandrine A (steroid with N -containing C-17 side-chain)
  - 8. Taxol (a modified diterpene pseudo alkaloid)
- II . Heterocyclic or typical alkaloids, divided into 14 groups according to their ring structure.
  - 1. Pyrrolidine alkaloids e.g., Hygrine;
  - 2. Piperidine alkaloids e.g., Lobeline;
  - 3. Pyrrolizidine alkaloids e.g., Senecionine;
  - 4. Tropane alkaloids e.g., Atropine;
  - 5. Quinoline alkaloids e.g., Quinine;
  - 6. Isoquinoline alkaloids e.g., Morphine;
  - 7. Aporphine alkaloids e.g., Boldine;
  - 8. Indole alkaloids e.g., Ergometrine;
  - 9. Imidazole alkaloids e.g., Pilocarpine;
  - 10. Diazocin alkaloids e.g., Lupanine;
  - 11. Purine alkaloids e.g., Caffeine;
  - 12. Steroidal alkaloids e.g., Solanidine;
  - 13. Amino alkaloids e.g., Ephedrine;
  - 14. Diterpene alkaloids e.g., Aconitine.

#### 1.3. Pyrrolizidine alkaloids

#### 1.3.1. Occurrence and distribution

The group of compounds known as pyrrolizidine alkaloids comprises over three hundred compounds identified to date, the majority of which occur naturally, although an increasing number are being derived synthetically or semi synthetically[22].

Naturally occurring pyrrolizidines are predominantly of plan origin and have been isolated from over four hundred and fifty species distributed among eighty-seven genera of fourteen plant families[23, 24]. The largest number of pyrrolizidine alkaloids occur in the genus Senecio. For this reason, and because the first pyrrolizidine alkaloid was isolated from this genus41, these alkaloids are often referred to as the Senecio alkaloids[22].

Pyrrolizidine alkaloids (Pyrrolizidine alkaloids) are a well-known class of defense compounds with a wide variety of structures. belong to the plant families Asteraceae, Boraginaceae, Orchidaceae and Fabaceae[25],

More than 660 pyrrolizidine alkaloids and N-oxide derivatives have been identified in over 6000 plants of previous families are considered to be potentially toxic[26]. The main sources of these toxic alkaloids are almost all genera of the family Boraginaceae, the tribes Senecioneae and Eupatorieae of the family Asteraceae and the genus Crotalaria of the family Fabaceae[27]

plants containing pyrrolizidine alkaloids are known to be toxic and have caused widespread livestock losses in many parts of the world[24]. Human poisoning has also been reported to occur through ingestion of pyrrolizidine alkaloids from contaminated foodstuffs and traditional herbal remedies and teas[28]. Some pyrrolizidine alkaloids have also been shown to exhibit anti-tumour activity and as a result are of great interest in medical research fields[28, 29].

Pyrrolizidine alkaloids have also been used as chemotaxonomic markers in various botanical studies[30, 31].

Pyrrolizidine alkaloids have also been isolated from some animal sources. Butterflies and moths of various genera feed on pyrrolizidine-containing plants and the ingested alkaloids are used for defence against predators and as pheromones[32, 33]. For example, the black and white moth Gnophaela latipennis (Arctiidae) has recently been shown to contain pyrrolizidines[34], whilst other workers have demonstrated the uptake of pyrrolizidines from host plants by the parasitic genus Pedicularis. In all these instances the pyrrolizidines were obtained from plant sources in the feeding process[35].

There is a group of naturally occurring Pyrrolizidine alkaloids which are not of plant origin; a number of ant genera have been shown to produce pyrrolizidines as defence compounds[36].

#### 1.3.2. Pyrrolizidine alkaloid Structure

All pyrrolizidine alkaloids contain the basic Pyrrolizidine ring figure (1). The molecules comprises two five-membered rings which share a nitrogen atom at position 4, this structural core is bicyclic ring system named systematically azabicyclo[3,3,0]octane(1). The diagram shows the traditional system for numbering the ring, which has been adopted throughout this work. The orientation of substituents is labelled as shown in figure (1). The fully saturated ring structure is non-planar and adopts an open 'V' shaped configuration. Whether the V opens towards or away from the viewer depends on the orientation of the substituents on C8[37, 38].

Most pyrrolizidine alkaloids identified so far have the basic structure system as shown in figure (1). A few alkaloids possess substituents such as methyl, methylene, carboxaldehyde, Carboxylic acid and nitrogen groups at C1, while some possess hydroxyl groups at other positions on the ring system, usually at C2 or C6. One or both of the hydroxyl groups (C7 or C9) may be esterified with a variety of acids[39]. The unesterified ring system is usually referred to as the amino alcohol "necine" and the acid portion as the "necic acid"[37, 38].

Pyrrolizidine alkaloids are esters of hydroxylated methyl pyrrolizidines, consisting of a necine base and necic acid moiety see figure (1). The necine base can either be 1,2-unsaturated (e.g. Retronecine (8) and Supinidine (12) or saturated (e.g. Alexine(3), Australine(4))[24]. The hepatotoxic pyrrolizidine alkaloids are monoesters, diesters or macrocyclicdiesters of unsaturated necine or otonecine bases[40].

Pyrrolizidine alkaloids can be oxygenated at N position to produce N-oxides. The polar nature of the N-oxide bond greatly enhances water solubility upon these alkaloids, relative to the corresponding tertiary bases[14]. Pyrrolizidine alkaloid N-oxides are unstable, they are easily converted into the tertiary amines in the presence of weak reducing agents[41]. To date, about 200 structures have been characterized, but many more have yet to be identified[42].

$$\begin{array}{c} R_2 \\ O \\ O \\ N \end{array} \begin{array}{c} O \\ R_1 \\ \text{necine base} \end{array}$$

Figure 1: The pyrrolizidine structure

#### 1.3.3. The Necine Bases and Simple alkaloids

There are over forty different necine bases, but only about twenty of these occur commonly[24]. Unesterified bases are rarely extracted from plants, but are obtained by hydrolysis of the alkaloid extract. Simple pyrrolizidine alkaloids are not esterified and are composed of a modified necine base.

The necine bases are hydroxyl derivatives of 1-methylpyrrolizidine compound[43], which consisting of two fused five-membered rings with bridgehead nitrogen. The fully saturated ring structure is non-planar and adopts an open 'V' shaped configuration Whether the V opens towards or away from the viewer depends on the orientation of the substituents on Carbon atoms[37, 38]. The necine bases are generally classified into five main types, Retronecine (8)

(RET), Heliotridine (9), crotanecine (10), supinidine(11), otonecine(12) (OTO)[43]. The necine can either be saturated or possess a double bond in the 1,2-position. Moreover, they may additionally bear one or two hydroxyl groups at C-2, C-6 or C-7 resulting in the formation of stereoisomers. Dashed and thickened (wedges) lines denote  $\alpha$  and  $\beta$ -orientations of bonds, respectively;  $\alpha$  meaning orientation away from the observer,  $\beta$  toward the observer[39].

The otonecine (12) is exception to the rest of the necines bases, in that it does not derived from 1-methylpyrrolizidine compound, but derived from an N-methylated azacyclooctan-4-one System. It act as a pyrrolizidine ring System by resonance operation leads to binding between N atom and the CO group[39]. The Pyrrolizidine alkaloids derived from these structures constitute a subgroup of the otonecine (12) alkaloids (O Pyrrolizidine alkaloids).

All known Pyrrolizidine alkaloids found in the plants can form N-oxide derivatives except the otonecine (12) alkaloids. The corresponding esterification of necines containing a double bond in the 1,2- position results in the formation of the toxic alkaloids[44].

A number of new and simple pyrrolizidine alkaloids compounds have recently been identified[22].

Figure 2: The structures of several necine bases and simple Pyrrolizidine alkaloids

#### 1.3.4. Necic acids

The acid Which is esterified with necine base called necic acid. Exception of acetic acid necic acids containing 4 to 10 carbon atoms and differ from each other in their composition. Where they may be mono or di-carboxylic acids with branched carbon chains. And containing substituted groups such as hydroxy, methoxy, epoxy, carboxy, acetoxy or other alkoxy groups[39]; and they can also be found in the form of Stereo- and diastereoisomers[44]. Figures (3 and 4) illustrate most important Necic acids that have been detected in alkaloids sofar.

The esterification possibilities are exemplified by several alkaloids. Necines containing one hydroxy group can be esterified with one monocarboxylic acid only as in Amabiline (40). Necines bearing two hydroxy groups such as 7,9-necinediols can be esterified with a

monocarboxylic acid either in the 7- or 9-position as demonstrated by7-angeloyl respectively 9-Angeloylretronecine (56). Echimidine (32) is an example of a twofold esterification. With dicarboxylic acids a double esterification takes place exclusively to form macrocyclic diesters (linking C7 with C9) of a necine [14, 45]. Base leading to the formation of alkaloids with 11- to 14-membered ring Systems. The most widely known Pyrrolizidine alkaloids are the11-membered monocrotaline, the 12-membered alkaloids senecionine and senkirkine, the 13-membered doronenine, and the 14-membered parsonsine.

Through combination of necines with necic acids an unimaginably large number of alkaloids may be theoretically obtained In nature.more than about 640 alkaloids were found so far and their structures elucidated[46].

#### 1.3.5. Monoester alkaloids

Monoester alkaloids are composed of a necine base esterified at either C7 or C9. A number of previously known monoester alkaloids have been isolated from new plant sources. Echinatine (50) and Lycopsamine (21)were isolated from *Anchusa arvensis*[47] Heliotrine (53)and Rinderine (47)were isolated from *Arnebia decumbens*[48].

#### 1.3.6. Acyclic diester alkaloids

Most new alkaloids of this class have Retronecine (8) as thenecine base.

Hydroxymyoscorpine (30)was isolated from *Llthospermwn erythrorhlzon*, along with Myoscorpine (29)andIntermedine (18)[49]. Doriasenine (49) was isolated from *Senecio doria*[50]

#### 1.3.7. Macrocyclic diester alkaloids

A macrocyclic diester alkaloid is composed of a necine base esterified to a dibasic acid, to produce a macrocyclic structure. Many such alkaloids are known [22].

Necic acids with two carbon atoms Acetic acid (Ac) Necic acids with five carbon atoms  $R_1$  $R_2$  $R_3$ Me Me Senecioic acid (Sen) Me Me Η Tiglic acid (Tig) Η Angelic acid (Ang) Me Sarracinic acid (Sar)  $CH_2OH$ Η Me Necic acids with seven carbon atoms  $R_2$  $R_3$ OH(-)-Viridifloric acid ОН Η (+)-Trachelanthic acid OHEchimidinic acid OHΗ Lasiocarpic acid OH Η OMe ОН (-)-Trachelanthic acid ОН (+)-Viridifloric acid Η

Figure 3: The Most Important Monocarboxylic Necic Acids

Necic acids with eight carbon atoms ОН Monocrotalic acid OHCrotaleschenic acid Necic acids with ten carbon atoms  $R_1$  $R_2$  $R_3$  $R_4$ Me Η Η Η Senecinic acid Η Me Η Η Integerrinecic acid Η Η Me Η Senecivernic acid Me OH Η Η Isatinecic acid Η Η OHMe Retronecic acid Me Η Η Seneciphyllic acid Η Me Η Spartioidinic acid Me Η OH Riddelliic acid Η Η Incanic acid Η OHTrichodesmic acid OHOHGlobiferic acid Erucifolinecic acid HO. Petasinecic acid

Figure 4: Most Important Dicarboxylic Necic Acids

#### 1.3.8. Biosynthesis of Pyrrolizidine Alkaloids

By using radioactive precursors it has been shown that pyrrolizidines are biosynthesized from the amino acid L-omithine via putrescine. Incorporation of two molecules of putrescine into homospermidine has been confirmed by enzymatic experiments. Homospennidine synthase catalyses the formation of homospermidine from two molecules of putrescine[51, 52].

An indication of how homospermidine is converted into necines was provided by the conversion of this triamine into trachelantharnidine using enzymes under physiological conditions [53, 54]. It is likely that oxidation of one terminal amino group of homospermidine produces an immonium ion [55], which can then undergo oxidation of the remaining primary amino group, followed by a non-enzymatic intramolecular cyclization of the aldehyde to give the more stable 1-formylpyrrolizidine. Reduction of this aldehyde then afforded trachelantharnidine. The formation of this base using enzymes suggested that the simple 1-hydroxymethylPyrrolizidine and trachelanthamidine should be tested as intermediates in the biosynthesis of more complex necines such as retronecine and heliotridine[55] as shows in scheme (1).

The biosynthetic pathways outlined in scheme (1) showed that the tracnelanthamidine is an efficient precursor for the retronecine and heliotridine; and the immonium ion is a key intermediate in the biosynthetic pathway of different necine bases including retronecine, heliotridine, rosmarinecine, isoretronecanol and supinidine from homospermidine in the plant species [56].

Otonecine, on the other hand, basic component of the otonecine alkaloids, is produced from retronecine, presumably by further hydroxylation and formation of a ketonic group with simultaneous cleavage of the C-N bond and N-methylation[57].

Scheme 1: Biosynthesis of different necines bases

#### 1.3.9. Analytical separation techniques

To determine pyrrolizidine alkaloid by analytical methods, the alkaloids must be present as free alkaloids. As pyrrolizidine alkaloid extracted from plants - with the exception of the otonecine alkaloids - generally occur as N-oxides and are thus only poorly soluble in organic solvents but readily soluble in water, the respective mixture should be reduced with zinc dust in the presence of dilute mineral acid[44]

In many cases, plants contain complex mixtures of alkaloids which cannot easily be separated, and the total alkaloid content of the plant is very small. Accordingly, pure components cannot be obtained in sufficient quantity for conventional analysis and other methods have to be used to analyze such mixtures[38].

Analytical thin-layer chromatography is a very useful technique. Silica coated plates are used most frequently for pyrrolizidine alkaloid work. Alkaloids are detected on chromatograms by a variety of spray reagents. Dragendorff's reagent is commonly used. The iodoplatinate reagent is more specific, but also more expensive. The most convenient and sensitive method for visualising unsaturated pyrrolizidine alkaloids is to spray the chromatogram with a chloranil solution, heating briefly, then spraying with Ehrlich's reagent[58].

Thin-layer chromatography can thus give an indication of the number and type of pyrrolizidines present in the crude alkaloid mixture [59-65]. Tentative identification of alkaloids by comparison of Rf values is possible if standards are available, but many alkaloids have identical Rf values and components are often missed due to incomplete separation.

High-performance liquid chromatography (HPLC) may be used in conjunction with mass spectrometry for identification of the components of complex alkaloidal mixtures[24]

A more frequently used technique is tandem gas chromatography mass spectrometry (GC-MS). The main drawback of gas chromatography is that underivatized alkaloids often decompose due to the high temperatures used for the separation. Many workers have analyzed alkaloid samples by derivitazation .The recent development of specially inert columns for polar components has facilitated the GC study of underivatized pyrrolizidines and many workers now use GC-MS for analysis of complex alkaloid mixtures[66]. The alkaloid fraction of *Echium plantagineum* L was analyzed by gas chromatography and mass spectrometry in different ionization modes[67]. Twelve pyrrolizidine alkaloids were characterized and identified by this method. Unfortunately, mass spectrometry is unable to distinguish between isomeric alkaloids, and results obtained can therefore be ambiguous. Reference standards are necessary for comparison of retention times and mass spectra in order to identify compounds unambiguously.

#### 1.3.10. Mass spectrometry of Pyrrolizidine alkaloids

Pyrrolizidine alkaloids exhibit characteristic peaks in their mass spectra. It is possible to obtain a great deal of information about the structure of a Pyrrolizidine alkaloid from electron impact mass spectrometry.

It is possible to identify the type of necine base present using diagnostic peaks. Saturated bases, such as platynecine (124) show intense ions at m/z 82 or m/z 83, together with peaks at m/z 97or m/z 113[24, 68]. The fragmentation pathway giving rise to these peaks is shown in Scheme (2). Unsaturated bases such as Retronecine (8) show intense ions at m/z 80 or 81 and 95 or 111[22, 24]. As shown in Scheme (3). Otonecine (12) type bases exhibit characteristic peaks at m/z94, 96 and 110 [22, 24] Scheme (4).

Scheme 2: A fragmentation pathway for Saturated necine bases

Scheme 3: A fragmentation pathway for Retronecine

**Scheme 4: Fragmentations pathway for otonecine** 

The croalbinecine (125) type base is saturated and possesses a hydroxyl group at C2 see page 61. Dehydration occurs readily and the mass spectrum can thus exhibit peaks characteristic

of both a saturated and unsaturated base[69]. It may also be difficult to identify the necine base of an alkaloid when it is esterified, since peak intensities are altered and the esterifying acid can give rise to peaks which are misleading; for example, the presence of angelic acid in Heliosupine (42) gives rise to a peak at m/z 83, as shown in Scheme (5) [22, 69]. In esterified necines account must be taken of a range of diagnostic peaks.

Scheme 5: Fragmentation pathway for heliosupine

It is possible to ascertain whether the base is monoesterified or diesterified, and whether the diester is acyclic or macrocyclic. C9-monoesters of Retronecine (8) exhibit an intense peak at m/z 138, C7-monoesters at m/z 137 and acyclic diesters at m/z 136 Scheme (6) [70-72]. The corresponding monoesters of platynecine (124) exhibit peaks at m/z two mass units higher; i . e at m/z 138, 139 or 140[22].

Macrocyclic diesters exhibit the characteristic "triads" of peaks; a group of peaks at m/z 80 and 81 with a second group at m/z 119, 120, 121 and a third at m/z 136, 137, 138indicate an unsaturated base alkaloid[24]. "Triads" of peaks atm/z 82 and 83; m/z 121, 122, 123 and m/z

138, 139 and 140 are exhibited by saturated base alkaloids [24]. The triad at m/z 119,120 and 121 (or two mass units higher) also appears in the spectra of acyclic diesters and monoester alkaloids[22].

Scheme 6: Fragmentation of monoesters and acyclic diesters

A great deal of information can be obtained from the relative intensities of peaks in the spectrum. The C9 ester bond is weaker than the C7 bond and will break more easily; hence ions corresponding to this cleavage will be more intense than ions corresponding to cleavage at the C7 linkage Scheme (7). In the case of acyclic diesters this enables one to ascertain the mode of ester attachment. In the case of macrocyclic diesters however, confusion can occur, since the ion corresponding to C9 cleavage can also be formed by cleavage at C7 with subsequent rearrangement[73]. This is shown in Scheme (8).

Scheme 7: The cleavage of C-7 and C-9 linkages of Pyrrolizidine diester

Scheme 8: Rearrangement of macrocyclic diesters

Rearrangements play a large role in the fragmentation of acyclic diester and monoester alkaloids. Scheme (9) shows two possible mechanisms for such a rearrangement [22].

Scheme 9: Possible mechanisms for rearrangement in acyclic diesters

These fragmentation pathways apply mainly to the Retronecine (8) and platynecine (124)111 type bases. The less common Pyrrolizidine bases exhibit other characteristic peaks. A base peak at m/z70 is considered by a number of authors to be indicative of a pyrrolizidine in which ring A is unsubstituted Clarynecine (126)[74, 75], as is the combination of a base peak at m/z 83 and a prominent peak at m/z 98[76]. A base peak at m/z 82 occurring with a major peak at m/z 113 is characteristic of a 1-hydroxymethylpyrrolizidine with an OH at C6 or C7 (e.g. 114)[76]. This combination is also considered characteristic of platynecine (124) type pyrrolizidines[77]. A base peak at m/z 83 is typical of the macronecine base (127)[78]. The otonecine base (12) shows characteristic fragments at m/z 168, 150, 122, 110 and 96 [79], while a base peak at m/z 124 is typical of 1-methylenepyrrolizidines (128)[80].

Other typical diagnostic peaks occur. A peak at m/z M-44(loss of CO<sub>2</sub>) is typical of all macrocyclic diesters. A peak at m/z M-17 is indicative of an OH group at C12 since this peak

is not observed in the spectra of alkaloids lacking this group[81] (figure (5) shows the numbering). A peak at m/z M-31shows that a methyl ester is present)[78]. A base peak at m/z M-31 is indicative of the loss of an exocyclic CH<sub>3</sub>OH group[68].

A strong peak at m/z 180 is typical of a 7-Acetylretronecinederivative [82]. Acyclic diesters of Retronecine (8) in which the ester at C7 is an acyl group show intense fragments at m/z180, 136 and 120[70]. A peak at m/z M-15 indicates the presence of a CH<sub>3</sub> group on the nitrogen atom[81]. A peak at m/z M-18 indicates the loss of H<sub>2</sub>O, which is considered to occur when there is an OH at C15 [81] (figure (5) shows the numbering). Other workers have found that a peak at M-18 occurred when there was a CH<sub>2</sub>OH group present at C12[83]. Acyclic diesters of Retronecine (8) or platynecine (124) which possess a C7-angeloyloxy (or tigloyloxy) group exhibit intense peaks at m/z 220, due to cleavage of the C9 bond[84] The same acid group at C9 leads toan intense peak at m/z 237. A peak at m/z 101 is due to protonated angelic, tiglic or senecioic acid [84]. A peak at m/z253 is indicative of a retronecine ester of hydroxysenecioic acid and one at m/z 235 indicates a retronecine ester of hydroxyangelic acid[50].

Although mass spectrometry can provide useful information concerning the overall structure of a pyrrolizidine alkaloid, relative stereochemistry cannot be determined, nor can geometric isomers always be distinguished. It is necessary to make use of reference standards for unambiguous structural assignment. Mass spectrometry is a powerful tool in its own right, but the information it provides can be greatly enhanced by the concurrent use of other spectroscopic techniques, where possible[22].

Figure 5: Numbering of macroesters pyrrolizidine alkaloids

# 1.4. Occurrence of pyrrolizidine alkaloids in Boraginaceae family

plant	pyrrolizidine alkaloids	referenc e
A 11	Triangularine (76)	[45 Q5
Alkana	7-Angeloylretronecine (55)	[45, 85,
trinctoria	Dihydroxytriangularine (105)	86]
	7-Angeloylretronecine (55)	
	9-Angeloylretronecine (56)	
	7-Tigloylretronecine (60)	
	7-Senecioylretronecine (57)	
	9-Senecioylretronecine (59)	
A. orientalis	7-Angeloyl-9-(hydroxypropenoyl)retronecine (66)	[47]
A. orientatis	7-Tigloyl-9-(hydroxypropenoyl) retronecine (68)	[47]
	7-Angeloyl-9-(2,3-dihydroxypropanoyl) retronecine (62)	
	7-Tigloyl-9-(2,3-dihydroxypropanoyl) retronecine (67)	
	Triangularine (76)	
	Triangularine (76)	
	Dihydroxytriangularine (105)	
	7-Angeloylretronecine (55)	
	7-Tigloylretronecine (60)	
	9-Tigloylretronecine (61)	
	Trachelanthamine (92)	
A tuboroulata	7-Angeloyl-9-(hydroxypropenoyl)retronecine (66)	[47]
A. tuberculata	7-Tigloyl-9-(hydroxypropenoyl) retronecine (68)	[47]
	7-Acetyl-9-sarracinoylretronecine (75)	
	7-Angeloyl-9-(2,3-dihydroxypropanoyl) retronecine (62)	
	7-Tigloyl-9-(2,3-dihydroxypropanoyl) retronecine (67)	
	Triangularine (76)	

	Triangularine (76)	
	Dihydroxytriangularine (105)	
	9-Acetyltrachelanthamidine (87)	
	9-Angeloyltrachelanthamidine (88)	
	Supinine (107)	
Anchusa	Intermedine (18)	
	Lycopsamine (21)	[47]
arvensis	7-Acetylintermedine (19)	
	7-Acetyllycopsamine (22)	
	7-diacetylintermedine (20)	
	7-Diacetyllycopsamine (23)	
A. arvensis	Echinatine (50)	[85]
	Retronecine(8)	
	Heliotridine (9)	
	7-Angeloylretronecine (55)	
A. milleri	Supinine (107)	[47]
A. milleri	Viridiflorine (114)	[47]
	Rinderine (47)	
	9-Curassavoylheliotridine (84)	
	7-Acetyl-9-curassavoylheliotridine (80)	
	Laburnine (93)	
	Acetyllaburnine (94)	
A. officinalis	Intermedine (18)	FAE 05
	Lycopsamine (21)	[45, 85,
	7-Acetyllycopsamine (22)	87-89]
	Clivorine (123)	
Borago	Intermedine (18)	
officinalis	Lycopsamine (21)	
	I and the second	Į.

	7-Acetylintermedine (19)	[45, 59-
	7-Acetyllycopsamine (22)	61, 85]
	Amabiline (40)	
	Supinine (107)	
	Thesinine (110)	
	Thesinine- 4'-O-β-D-glucoside (111)	
	no Pyrrolizidine alkaloids could be detected in extracts of	
Cordia gilletii.	the root bark and leaves of the Congolese specimen of	[90]
	Cordia gilletii.	
	Supinine (107)	
Cymaglaggym	Amabiline (40)	
Cynoglossum amabile	Rinderine (47)	[91]
атавне	Echinatine (50)	
	7-Acetylechinatine (51)	
	Heliosupine (42)	
	12-Acetylheliosupine (44)	
	Echinatine (50)	
	7-Angeloylheliotridine (79)	
	Viridiflorine (114)	
C. officinale	7-Tigloylheliotridine (83)	
C. Officinale	7-Angeloyl-1-fomryl-6,7-dihydro-5H-pyrrolizdine (121)7-	[39, 45,
	Angeloylrinderine (48)	91, 92]
	Rinderine (47)	
	7-Angeloyl-9- (2-methylbutyryl) heliotridine (81)	
	7-Angeloyl-9- (2.3-dihydroxybutyryl) heliotridine (81)	
	7-Angeloylechinatine (52)	
	Echinatine (50)	
	Echinatine (50)	

	7-Tigloylheliotridine (83)	
	Rinderine (47)	
	7-Angeloylrinderine (48)	
	7-Angeloyl-1-fomryl-6,7-dihydro-5H-pyrrolizdine (121)	
	7-Angeloyl-9- (2-methylbutyryl) heliotridine (81)	
	7-Angeloyl-9- (2.3-dihydroxybutyryl) heliotridine (81)	
E 1:	Echimidine (32)	
Echium	7-Angeloylretronecine (55)	[62, 85]
amoenum	7-Tigloylretronecine (60)	
	(7S, 8R)-Petranine (115)	
	(7S, 8S)-petranine (116)	
П 1	(7R, 8R)-petranine (117)	F45 021
E. glomeratum	(7R, 8S)-petranine (118)	[45, 93]
	7-Angeloylretronecine (55)	
	9-Angeloylretronecine (56)	
	7-Angeloylretronecine (55)	
	7-Tigloylretronecine (60)	
	Lycopsamine (21)	
	7-Angeloyl -9-(2-methylbutyryl)retronecine (63)	
	7-Tigloyl-9-(2-methylbutyryl) retronecine (69)	
	7-Acetyllycopsamine (22)	FO 41
E. Horridum	Uplandicine (26)	[94]
	7-Angeloyl-9-(2,3-dihydroxybutyryl)retronecine (65)	
	7-Tigloyl-9-(2,3-dihydroxylbutyryl)retronecine (70)	
	7-Angeloyllycopsamine (25)	
	7-Tigloyllycopsamine (24)	
	Echimidine (32)	
E. pininana	Myoscorpine (29)and its N-oxide	[95]

	Hydroxymyoscorpine (30)	
	Echimidine (32	
	7-Acetylintermedine (19)	
	Echiupinine (58) and its N-oxide	
	7-Angeloylretronecine (55)	
	7-Tigloylretronecine (60)	
	Lycopsamine (21)	
	7-Angeloyl -9-(2-methylbutyryl)retronecine (63)	
	7-Tigloyl-9-(2-methylbutyryl) retronecine (69)	
T. 10	7-Acetyllycopsamine (22)	10.41
E. rauwolfii	Uplandicine (26)	[94]
	7-Angeloyl-9-(2,3-dihydroxybutyryl)retronecine (65)	
	7-Tigloyl-9-(2,3-dihydroxylbutyryl)retronecine (70)	
	7-Angeloyllycopsamine (25)	
	7-Tigloyllycopsamine (24)	
	Echimidine (32)	
	Echimidine (32)	
	7-Angeloylretronecine (55)	
	9-Angeloylretronecine (56)	
	7-Tigloylretronecine (60)	
E	9-Tigloylretronecine (61)	1061
E. setosum	7-Angeloyl -9-(2-methylbutyryl)retronecine (63)	[96]
	7-Tigloyl-9-(2-methylbutyryl) retronecine (69)	
	Uplandicine (26)	
	7-Angeloyl-9-(2.3-dimelhylbutyryl)retronecine (64)	
	7-Angeloyl-9-(2,3-dihydroxybutyryl)retronecine (65)	
E sudo ser	7-Tigloylretronecine (60)	[85, 96,
E. vulgare	9-Tigloylretronecine (61)	97]

	7-Angeloyl -9-(2-methylbutyryl)retronecine (63)	
	7-Angeloyl-9-(2,3-dihydroxybutyryl)retronecine (65)	
	7-acetylvulgarine (14)and its N-oxide	
	7-Tigloyl-9-(2-methylbutyryl) retronecine (69)	
	9-Angeloylretronecine (56)	
	9-Senecioylretronecine (59)	
	Acetylechimidine (34) and its N-oxide	
	Echimidine (32) and its N-oxide	
	Echimiplatine (17)	
	Echiuplatine (122)	
	7-Echivulgarine (15) and its N-oxide	
	Hydroxymyoscorpine (30)	
	Leptanthine (16) and its N-oxide	
	Retronecine (8)	
	Uplandicine (26) and its N-oxide	
	Vulgarine (13) and its N-oxide	
E. wildpretti	Echimidine (32) and its N-oxide	[63]
E. angustifolium	Echimidine (32)	[98]
E. diffusum	Heliotridine (9)	[99]
	Echimidine (32)	
	Echihumiline (35)	54.007
E. humile	Lycopsamine (21)	[100]
	7-Acetyllycopsamine (22)	
	Echimidine (32),	
E.	Echihumiline (35)	F1013
hypertropicum	7-Senecioylretronecine (57)	[101]
	9-Angeloylretronecine (56)	

	Lycopsamine (21)	
E. italicum	Echimidine (32)	[99]
	Echimidine (32)	
	Echiumine (102)	
	Uplandicine (26)	
E. lycopsis	Lycopsamine (21)	[97, 10
	Intermedine (18)	
	Echiuplatine (122)	
	9-Angeloylretronecine (56), Leptanthine (16).	
. ·	Echimidine (32)	[102]
E. sericeum	Symlandine (27) (26) (or Symphytine (31)).	[103]
	7-Angeloylretronecine (55)	F1041
E. simplex	9-Angeloylretronecine (56).	[104]
E. stenosiphon		
Webb subsp.	Echimidine (32)	[101]
stenosiphon		
E. tuberculatum	Echimidine (32)	[99]
Ehretia aspera	Ehretinine (112)	[99]
Eritrichium	7. A	[00]
rupestre	7-Angeloylretronecine (55)	[99]
H. longituba	9-Angeloylretronecine (56).	[99]
Heliotropium	11 1: 4: (52)	[105]
acutifolium	Heliotrine (53)	[105]
H. amplexicaule	Indicine (99)	[99]
H.	Retronecine (8)	5106
angiospermum	Supinidine (11)	[106]
	Europine (108)	500 11
H. arbainense	Heliotrine (53)	[99, 10

	Lasiocarpine (45).	
H. arborescens	Indicine (99)	[00]
	Lasiocarpine (45)	[99]
H. arguzioides	Heliotrine (53)	[99]
	Europine (108)	
H. bacciferum	Heliotrine (53)	[00]
n. vaccijerum	Heleurine (106)	[99]
	Supinine (107)	
-	Europine (108)	
II banai	7-Acetyleuropine (109)	1001
H. bovei	Lasiocarpine (45)	[99]
	Lasiocarpine (45) N-oxide	
H. bracteatum	Helibractinecine (89)	[99]
11. Dracteatum	Retronecine (8)	[99]
H. bursiferum	7-Angeloylretronecine (55).	[99]
	Echinatine (50)	
	Europine (108)	
H. circinatum	Heleurine (106)	[99]
	Heliotrine (53)	
	Lasiocarpine (45)	
H.	Retronecine (8)	[106]
confertifolium	Supinidine (11)	[106]
H. crassifolium	Europine (108) and its N-oxide	[99]
Н.	7-Angeloylheliotridine (79)	[106,
	Retronecine (8)	108]
curassavicum	Supinidine (11)	100]
H. dasycarpum	Heliotrine (53)	[99]

H. digynum (H.	Europine (108)	
	Heliotrine (53)	[99]
luteum)	Lasiocarpine (45)	
	Heliotrine (53)	
H. disciforme	Heliotrine (53) N-oxide	[99]
	Heleurine (106)	
	Heliotrine (53)	
H. dissitiflorum	Heliotrine (53) N-oxide	[99]
	Europine (108).	
	Heliotrine (53)	
H. eichwaldii	7-angeloylheliotrine	[99]
	Lasiocarpine (45).	
H osfandianii	Europine (108)	[100]
H. esfandiarii	Europine (108) N-oxide.	[109]
	Europine (108)	
	Heleurine (106)	
U aurongaum	Heliotrine (53)	[109,
H. europaeum	Lasiocarpine (45)	110]
	Heliotrine (53) n-oxide	
	Supinine (107)	
H foliosisimum	Retronecine (8)	[106]
H. foliosisimum	Supinidine (11)	[100]
H. fruticosum	Retronecine (8)	[106]
11. jruncosum	Supinidine (11)	[100]
	Europine (108)	
H.	Heliotrine (53)	[108,
hirsutissimum	Heleurine (106)	111-113]
	Lasiocarpine (45)	
	1	1

	3'-Acetyllasiocarpine (46)	
	Supinine (107)	
-	Echinatine (50)	
	Helindicine (119)	
	Heliotrine (53)	
	Heleurine (106)	
	Indicine (99)	Γ1 <b>Λ</b> 0
H. indicum	Lasiocarpine (45)	[108,
H. inaicum	Lycopsamine (21)	111, 112, 114]
	Rinderine (47)	114]
	Supinine (107)	
	Retronecine (8)	
	Supinidine (11)	
	Trachelanthamine (92)	
H. keralense	Intermedine (18)	[115]
11. keruiense	Retronecine (8)	[113]
H. maris mortui	Europine (108)	[107]
11. marts mortat	Lasiocarpine (45).	[107]
H.	Lycopsamine (21)	[116]
megalanthum	Zyeopoulinie (21)	[110]
H. olgae	Heliotrine (53)	[117]
H. ovalifolium	Retronecine (8)	[118]
H. procumbens	Retronecine (8)	[106]
п. procumbens	Supinidine (11)	[100]
Н.	Retronecine (8)	[106]
queretaroanum	Supinidine (11)	[100]
H. racemosum	Retronecine (8)	[106]
11. racemosunt	Supinidine (11)	[100]
	•	•

Н.	Europine (108)	
	Heliotrine (53)	[70, 119]
rotundifolium	Lasiocarpine (45).	
H. scabrum	Heliscabine (91)	[120]
H. SCabrum	Retronecine (8).	[120]
U gagaj	Retronecine (8)	[106]
H. sessei	Supinidine (11).	[106]
	Amabiline (40)	
H. spathulatum	Retronecine (8)	[23, 108]
	Supinidine (11)	
H. steudneri	Lycopsamine (21)	[24]
H. strigosum	Strigosine (113)	[121]
	Retronecine (8)	
H. subulatum	Heliotrine (53)	[122]
	7-Angeloylheliotridine (79)	
	Echinatine (50)	
	Heliosupine (42)	
H. supinum	Heliotrine (53)	[24]
11. зариши	7-Angeloylheliotridine (79)	[24]
	Lasiocarpine (45)	
	Supinine (107).	
	Intermedine (18)	
	Indicine (99)	
H. transalpinum	Lycopsamine (21)	[123]
	Rinderine (47)	
	Supinine (107).	
H. transoxanum	Heliotrine (53)	[105]

H. wigginsii	Retronecine (8)	[106]
	Supinidine (11)	[100]
	Retronecine (8)	
	7-Angeloylheliotridine (79)	
	Trachelanthamine (92)	
Gastrocotyle	Supinine (107)	[47]
hispida	Trachelanthamine (92) isomer	[47]
	Intermedine (18)	
	Lycopsamine	
	7-Acetyllycopsamine (22)	
Heliotropium	Indicine (99)	[85, 124]
amplexicaule	marcine (33)	[65, 124]
	Lasiocarpine (45)	
H. arborescens	Indicine (99)	[39]
	12-Acetylindicine (101)	
H.hovei	7-Acetyleuropine (109)	[61]
	Strigosine (113)	
	Supinine (107)	
	Heleurine (106)	
	Heliotrine (53	
	Indicine (99)	
H. indicum L.	3-Acetylindicine (100)	[124,
H. maicum L.	Lycopsamine (21)	125]
	Echinatine (50)	
	Europine (108)	
	Heliosupine (42)	
	Lasiocarpine (45)	
	Helindicine (119)	

H. rotundifolium	Europine (108) N-oxide	[119]
	Retronecine (8)	
H. scabrum	Helifoline (90)	[120]
	Helibractinecine (89)	
H. arborescens	Indicine (99)	[45]
11. urborescens	12-Acetylindicine (101)	[40]
	Heliotrine (53)	
	Europine (108)	
H. europaeum	Lasiocarpine (45)	[45]
	Supinine (107)	
	Heleurine (106)	
	Indicine (99)	
	eliotrine (53)	
	Supinine (107)	
	Lasiocarpine (45)	
II : 1:	Echinatine (50)	
H. indicum	Europine (108)	[45]
	Heleurine (106)	
	Helindicine (119)	
	Heliosupine (42)	
	Lycopsamine (21)	
Lappula intermedia	Lasiocarpine (45)	[125]
	7-Acetyl-9-sarracinoylretronecine (75)	
L. spinocarpos	7-Angeloylheliotridine (79)	[47]
L. spinocurpos	7-Acetylintermedine (19)	[47]
	7-Acetyllycopsamine (22)	
	ı	T.

	Amabiline (40)	
	7-Angeloylheliotridine (79)	
	Intermedine (18	
	Lycopsamine (21)	
	Retronecine (8)	
	Supinine (107)	
	Trachelanthamine (92)	
	Viridiflorine (114)	
	Lycopsamine (21)	
L. myosotis	7-Acetyllycopsamine (22)	[126]
Moench	Intermedine	[120]
	7-Acetylintermedine (19)	
	Lycopsamine (21)	
	7-Acetyllycopsamine (22)	
Lithosparmum	7-Acetylintermedine (19)	
Lithospermum	Canescine (95)	[127]
canescens	Canescenine (97)	
	13-Acetylcanescine (96)	
	13-acetylcanescenine (98)	
L. erythrorhizon	Intermedine (18)	[128]
I officianale	Lithosenine (103)	[39, 45,
L. officinale	12-Acetyllithosenine (104)	61, 85]
Maris-Mortui	Europine (108)N-oxide	[119]
Myosotis	Myoscorpine (29)	
•	7-Acetylscorpioidine (39)	[85]
palustris	Symphytine (31)	
M. saarniaidas	Myoscorpine (29)	[20, 45]
M. scorpioides	Scorpioidine (38)	[39, 45]
	ı	

	7-Acetylscorpioidine (39) (38)	
	Symphytine (31)	
	Intermedine (18)	
Onosma	Lycopsamine (21)	[120]
alborosea	7-Acetylintermedine (19)	[129]
	7-Acetyllycopsamine (22)	
	5,6-Dihydro-7,9-dimethoxy -7H- pyrrolizine (120)	
	7-Acetylretronecine (54)	
	9-(Butyryl-2-ene) supinidine (86) (85)	
	7-Acetyl-9-(2-melhylbutyryl) retronecine (72)	
	7-Acetyl-9-(2.3-dimelhylbutyryl)retronecine (71)	[120]
O. arenaria	7-Acetyl-9-(2-hydroxy-3-methylbutyryl) retronecine (73)	[130]
	3'-Acetylsupinine (41)	
	7-Acetyl-9-(2,3-dihydroxybutyryl)retronecine (74)	
	7-Acetyllycopsamine (22)	
	Uplandicine (26)	
	Echihumiline (35) and its N-oxide	
O. leptantha	Leptanthine (16)	[45]
	3'-Acetylechihumiline (36)	
	9-Angeloylretronecine (56)	
	Echimidine (32)	
O. alboroseum	Lycopsamine (21)	[131,
sanguinolentum	Intermedine (18)	132]
	7-Acetylintermedine (19)	
	7-Acetyllycopsamine (22).	
-	7-Acetyllycopsamine (22)	
O. arenaria	7-Acetylretronecine (54)	[130]
		1

	3'-Acetylsupinine (41)	
	Uplandicine (26).	
	Intermedine (18)	
O. arenaria	Lycopsamine (21)	[121]
subsp. pennina	7-Acetylintermedine (19)	[131]
	7-Acetyllycopsamine (22).	
	Echihumiline (35)	
O. leptantha	3'-Acetylechihumiline (36)	[133]
	Leptanthine (16) and their N-oxides.	
	Echimidine (32)	
	Leptanthine (16)	[124]
O. stellulatum	Lycopsamine (21) and heir N-oxides	[134]
	7-Acetylintermedine (19)	
	Uplandicine (26)	
	Heliotridine (9)	
	7-Angeloylrinderine (48)	
Paracaryum	7-Senecioylheliotridine (82)	[// <del>7</del> ]
intermedium	Supinine (107)	[47]
	Viridiflorine (114)	
	Rinderine (47)	
	7-Angeloylheliotridine (79)	
	Viridiflorine (114)	
Вамасатум	Rinderine (47)	
Paracaryum	Echinatine (50)	[47]
rugulosum	7-Angeloylrinderine (48)	
	7-Senecioylrinderine (49)	
	Heliosupine (42)	
	I .	I

	Asperumine (78)		
	Echiumine (102)		
Carreera la actuaria	Symlandine (27)		
Symphytum	Symphytine (31)	[85]	
asperum	Myoscorpine (29)		
	Echinatine (50)		
	Echimidine (32)		
	Lasiocarpine (45)		
	Intermedine (18)		
	Echimidine (32)		
	Lycopsamine (21)		
C	Symlandine (27)	[39, 45]	
Symphytum	Symviridine (28)		
asperum	Heliosupine (42) N-oxide		
	Myoscorpine (29)		
	Echinatine (50)		
	Symphytine (31)		
	Asperumine (78)		
	Echimidine (32)		
Symphytum	Asperumine (78)	F457	
caucasicum	Echinatine (50)	[45]	
	Lasiocarpine (45)		
Caranhartura	Echimidine (32)		
Symphytum	Asperumine (78)	[20]	
caucasicum	Echinatine (50)	[39]	
Bieb	Lasiocarpine (45)		
	1	I	

	7-Acetylintermedine (19)	
	7-Acetyllycopsamine (22)	
	Echimidine (32)	
	Echinatine (50)	
	Heliosupine (42)	
	Heliotrine (53)	120 45
Symphytum	Intermedine (18)	[39, 45,
officinale	Lasiocarpine (45)	85, 124,
	Lycopsamine (21)	135]
	Myoscorpine (29)	
	Symlandine (27)	
	Symphytine (31)	
	Symviridine (28)	
	Viridiflorine (114)	
G 1 .		
Symphytum	Echimidine (32) and its Novide	[136]
sylvaticum	Echimidine (32) and its N-oxide	[136]
	Echimidine (32) and its N-oxide  Symlandine (27)	[136]
		[136]
sylvaticum	Symlandine (27)	
Symphytum	Symlandine (27) Echimidine (32)	[39, 45,
Symphytum	Symlandine (27) Echimidine (32) Anadoline(37)	[39, 45,
Symphytum tuberosum L	Symlandine (27) Echimidine (32) Anadoline(37) 7-Acetyllycopsamine (22)	[39, 45,
Symphytum tuberosum L  Symphytum	Symlandine (27) Echimidine (32) Anadoline(37) 7-Acetyllycopsamine (22) Intermedine (18)	[39, 45,
Symphytum tuberosum L	Symlandine (27) Echimidine (32) Anadoline(37) 7-Acetyllycopsamine (22) Intermedine (18) Lycopsamine (21)	[39, 45, 85]
Symphytum tuberosum L  Symphytum	Symlandine (27) Echimidine (32) Anadoline(37) 7-Acetyllycopsamine (22) Intermedine (18) Lycopsamine (21) Symlandine (27)	[39, 45, 85]
Symphytum tuberosum L  Symphytum	Symlandine (27) Echimidine (32) Anadoline(37) 7-Acetyllycopsamine (22) Intermedine (18) Lycopsamine (21) Symlandine (27) Symphytine (31)	[39, 45, 85]
Symphytum tuberosum L  Symphytum uplandicum	Symlandine (27) Echimidine (32) Anadoline(37) 7-Acetyllycopsamine (22) Intermedine (18) Lycopsamine (21) Symlandine (27) Symphytine (31) Uplandicine (26)	[39, 45, 85]
Symphytum tuberosum L  Symphytum uplandicum  Symphytum x	Symlandine (27) Echimidine (32) Anadoline(37) 7-Acetyllycopsamine (22) Intermedine (18) Lycopsamine (21) Symlandine (27) Symphytine (31) Uplandicine (26) Intermedine (18)	[39, 45, 85] [45]

	7-Acetyllycopsamine (22)	
	Uplandicine (26)	
	Symlandine (27)	
	Symviridine (28)	
	Myoscorpine (29)	
	Symphytine (31)	
	Echimidine (32)	
Tournefortia	Supinine (107)	[124]
acuminata	Suprime (107)	[124]
	Retronecine (8)	
Trichodesma	Viridiflorine (114)	[4 <b>7</b> ]
africanum	Intermedine (18)	[47]
	Lycopsamine (21)	
Trichodesma	Supinine (107)	[124]
zeylanicum	Suprimite (107)	[14 <sup>-</sup> 7]

$$\begin{array}{c} R_2 \\ R_5 \\ R_5 \\ \end{array}$$

$R_1$	$R_2$	$\mathbb{R}_3$	$R_4$	$R_5$		
ОН	Н	OAng	ОН	Н	Vulgarine	13
ОН	Н	OAng	OAc	Н	7-acetylvulgarine	14
ОН	Н	ОН	OAng	Н	Echivulgarine	15
ОН	Н	ОН	ОН	Н	Leptanthine	16
H	OH	ОН	ОН	Н	Echimiplatine	17
H	OH	Н	ОН	Н	Intermedine	18
H	OH	Н	OAc	Н	7-Acetylintermedine	19
H	OH	Н	OAc	OAc	7-diacetylintermedine	20
ОН	Н	Н	ОН	Н	Lycopsamine	21
ОН	Н	Н	OAc	Н	7-Acetyllycopsamine	22
ОН	Н	Н	OAc	OAc	7-Diacetyllycopsamine	23
ОН	Н	Н	OTig	Н	7-Tigloyllycopsamine	24
ОН	Н	Н	OAng	Н	7-Angeloyllycopsamine	25
ОН	Н	ОН	OAc	Н	Uplandicine	26
ОН	Н	Н	OAng	Н	Symlandine	27
ОН	Н	Н	OSen	Н	Symviridine	28
H	OH	Н	OTig	Н	Myoscorpine	29
H	ОН	ОН	OTig	Н	Hydroxymyoscorpine	30
ОН	Н	Н	OTig	Н	Symphytine	31
Н	OH	ОН	OAng	Н	Echimidine	32
Н	ОН	ОН	OTig	Н	Echimidine (isomer)	33
Н	ОН	OAc	OAng	Н	Acetylechimidine	34

Н	OH	ОН	OSen	Н	Echihumiline	35
Н	OAc	ОН	OSen	Н	3'-Acetylechihumiline	36
Н	OTig	Н	ОН	Н	Anadoline	37
OTig	Н	Н	ОН	Н	Scorpioidine	38
OTig	Н	Н	OAc	Н	7-Acetylscorpioidine	39
ОН	Н	Н	Н	Н	Amabiline	40
Н	OAc	Н	Н	Н	3' -Acetylsupinine	41
ОН	Н	ОН	Н	OAng	Heliosupine	42
OAc	Н	ОН	Н	OAng	3'-Acetylheliosupine	43
OAc	Н	ОН	Н	OAng	12-Acetylheliosupine	44
Н	OMe	ОН	Н	OAng	Lasiocarpine	45
Н	OAc	ОН	Н	OAng	3'-Acetyllasiocarpine	46
Н	ОН	Н	Н	ОН	Rinderine	47
Н	ОН	Н	Н	OAng	7-Angeloylrinderine	48
Н	ОН	Н	Н	OSen	7-Senecioylrinderine	49
ОН	Н	Н	Н	ОН	Echinatine	50
ОН	Н	Н	Н	OAc	7-Acetylechinatine	51
ОН	Н	Н	Н	OAng	7-Angeloylechinatine	52
Н	OMe	Н	Н	ОН	Heliotrine	53

$R_1$	$R_2$		
H	ОН	Retronecine	8
Н	OAc	7-Acetylretronecine	54
Н	OAng	7-Angeloylretronecine	55
Ang	ОН	9-Angeloylretronecine	56

Н	OSen	7-Senecioylretronecine	57	
(+)-Trac	OSen	Echiupinine	58	
Sen	ОН	9-Senecioylretronecine	59	
Н	OTig	7-Tigloylretronecine	60	
Tig	ОН	9-Tigloylretronecine	61	
2,3-dihydroxypropanoyl	OAng	7-Angeloyl-9-(2,3-dihydroxypropanoyl)	62	
		retronecine		
2-methylbutyryl	OAng	7-Angeloyl -9-(2-methylbutyryl)retronecine	63	
2.3-dimelhylbutyryl	OAng	7-Angeloyl-9-(2.3-dimelhylbutyryl)retronecine	64	
2.2 dibydrovyhutyryl	OAna	7-Angeloyl-9-(2,3-	65	
2,3-dihydroxybutyryl	OAng	dihydroxybutyryl)retronecine	03	
hydroxypropenoyl	OAng	7-Angeloyl-9-(hydroxypropenoyl)retronecine	66	
2.2.11.1	OTT'	7-Tigloyl-9-(2,3-dihydroxypropanoyl)	<b>67</b>	
2,3-dihydroxy propanoyl	OTig	retronecine	67	
hydroxypropenoyl	OTig	7-Tigloyl-9-(hydroxypropenoyl) retronecine	68	
2-methylbutyryl	OTig	7-Tigloyl-9-(2-methylbutyryl) retronecine	69	
2,3-dihydroxy propanoyl	OTig	7-Tigloyl-9-(2,3-dihydroxylbutyryl)retronecine	70	
2.3-dimelhylbutyryl	OAc	7-Acetyl-9-(2.3-dimelhylbutyryl)retronecine	71	
2-melhylbutyryl	OAc	7-Acetyl-9-(2-melhylbutyryl) retronecine	72	
2 hardwayy 2 mathailbutawy	040	7-Acetyl-9-(2-hydroxy-3-methylbutyryl)	73	
2-hydroxy-3-methylbutyryl	OAc	retronecine	13	
2,3-dihydroxybutyryl	OAc	7-Acetyl-9-(2,3-dihydroxybutyryl)retronecine	74	
sarr	OAc	7-Acetyl-9-sarracinoylretronecine	75	
t Sarr	OAng	Triangularine	76	
t Sarr	OTig	Triangularicine	77	
Tig	OTig	Asperumine	78	

$$R_2$$
 H  $OR_1$ 

$R_1$	$R_2$		
Н	ОН	Heliotridine	9
Н	OAng	7-Angeloylheliotridine	79
Cura	ОН	7-Acetyl-9-curassavoylheliotridine	80
2.3-dihydroxybutyryl	OAng	7-Angeloyl-9- (2.3-dihydroxybutyryl) heliotridine	81
2-methylbutyryl	OAng	7-Angeloyl-9- (2-methylbutyryl) heliotridine	82
Н	OSen	7-Senecioylheliotridine	83
Н	OTig	7-Tigloylheliotridine	84
Cura	ОН	9-Curassavoylheliotridine	85
Butyryl-2-ene	Н	9-(Butyryl-2-ene) supinidine	86
Ac	Н	9-Acetyltrachelanthamidine	87
Ang	Н	9-Angeloyltrachelanthamidine	88

$\mathbf{R}_1$	$R_2$	$\mathbb{R}_3$	$\mathbb{R}_4$		
Н	OH	ОН	ОН	Helibractinecine	89
ОН	ОН	Н	ОН	Helifoline	90
Н	OH	ОН	ОН	Heliscabine	91
Н	O(+)Trach	Н	Н	Trachelanthamine	92
Н	Н	CH <sub>2</sub> OH	Н	Laburnine	93
Н	Н	CH <sub>2</sub> OMe	Н	Acetyllaburnine	94

$$\begin{array}{c|c} & & & & \\ & &$$

 $R_1$  $R_2$ Н OHCanescine 95 Н 13-Acetylcanescine 96 OAc ОН Canescenine 97 Η 13-Acetylcanescenine OAc Н 98

 $R_1$  $R_2$ Indicine OHOH 99 3-Acetylindicine OH OAc 100 12-Acetylindicine OAc ОН 101 Echiumine 102 ОН OAng

R

H Lithosenine 103

OAc 12-Acetyllithosenine 104

Dihydroxytriangularine 105

$$HO \longrightarrow O \longrightarrow H$$

Thesinine 110

Thesinine- 4'-O-β-D-glucoside 111

Ehretinine 112

Strigosine 113

Viridiflorine 114

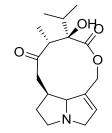
(7S, 8R)-Petranine 115

(7S, 8S)-Petranine 116

(7R, 8R)-Petranine 117

(7R, 8S)-Petranine

118



Helindicine 119

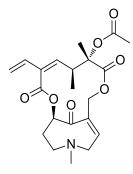
5,6-Dihydro-7,9-dimethoxy -7H- pyrrolizine 120

7-Angeloyl-1-fomryl-6,7-dihydro-5H-pyrrolizdine 121

$$\begin{array}{c|c}
O & H & O \\
\hline
O & N & O
\end{array}$$

$$\begin{array}{c|c}
O & (C_6H_{13}O_2) \\
O & O
\end{array}$$

Echiuplatine 122



Clivorine 123

macronecine 127 1-methylenepyrrolizidines 128

## 1.5. Aim of the study

The present study aimed to isolate and identify the pyrrolizidine alkaloid content of *Echium plantagineum* L which grown in Libya (El-marj) by the Retention index and Gas chromatography Mass Spectroscopy application . while the extraction and isolation is to be performed by acid base extraction technique of reduced N-oxide pyrrolizidine alkaloids and free alkaloids.

Proposal skeletons of isolated compound would be studied by comparison their mass fragmentations and retention indices with those of previously worked pyrrolizidine alkaloids as standard values.

#### **CHAPTER TWO**

#### 2. EXPERIMENTAL

## 2.1. The description of *Echium plantagineum* L.

Echium plantagineum L shown in figure (6) is an erect annual or biennial herb, 20-60 cm tall. With 1-many flowering stems. Covered with soft tubercle-based spreading hairs. Basal leaves usually in a rosette,  $\pm$  ovate.  $40\text{-}120 \times 12\text{-}15$  mm, with distinct, lateral nerves, covered with more or less soft appressed setae; cauline leaves smaller, more or less cordate at the base, obLong to lanceolate. Cyme terminal or axillary. Calyx 6-10 mm at anthesis. up to 15 mm in fruit, lobes linearlanceolate. Corolla blue, infundibuliform. 18-35 mm long; glabrous except on the veins and the margins; limb rounded, 11-14 mm in diameter. Only 2 stamens exserted; anthers violet, oblong. c. 1 mm long; style bifid, very sparcely pilose. Style bifid for nearly 1 mm. Nutlets 4, c.  $3\times3$  mm, ovoid, bipyramidal with dorsal and ventral keel, tuberculate and faintly striate[137].

#### 2.2. Plant Collection

Flowers, leaves, stems of *Echium plantagineum* L plant materials were collected from Elmarj, Gabel Akhder at the east Northern part of Libya during April 2012. Plant raw material spread in three square meter shaded area for air-drying at room temperature for about two weeks. After drying, exact weights were recorded was 378 gram. Whole plant grounded to powder using a laboratory hammer mill. Powdered materials were maintained at room temperature (25 °C), protected from light by storing the powder in dark glass container to avoid the effect of light on pyrrlolizidine alkaloids until required for extraction and analysis.

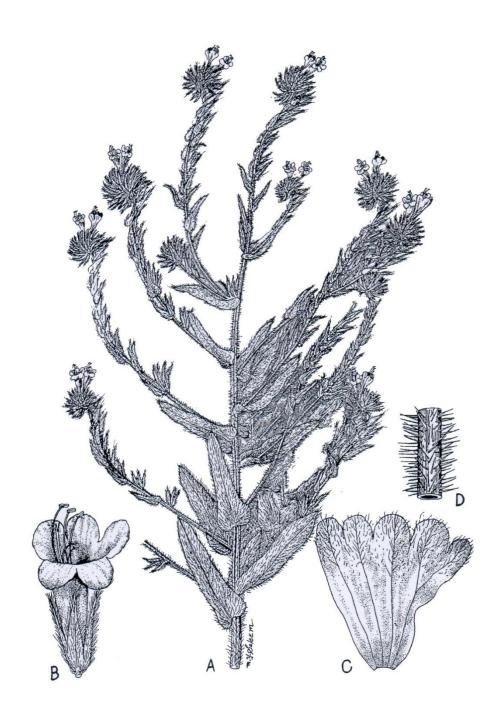


Figure 6: A. apportion of flowering stem $\times$ 0.5, B.flower $\times$ 2, C. dissected corolla $\times$ 2apportion of stem $\times$ 0.5

## 2.3. Solvent Extraction of *Echium plantagineum* L

Flowers, leaves, stems of dried plant materials (378 gram) were bowered down and macerate for 24 hours with 2L methanol with intermittent shaking.

The maceration process was then repeated several times for exhaustive extraction. The methanolic extract is then collected and concentrated almost to dryness by using rotary evaporator under vacuum at 40 °C[14].

## 2.4. Screening for alkaloids

The alkaloids nucleus could be detected by Dragendorff's reagent. The methanolic extract was dropped into a porcelain basin, dried by blower and added with a few drops of Dragendorff's reagent. A change occurred with red-brown precipitations within several minutes indicated the presence of alkaloid nucleus[138].

#### 2.5. Alkaloid extraction

All alkaloids contain at least one nitrogen atom, in the majority of cases, those compounds are basic. This means that salt formation can occur in the presence of acid. This fundamental property of alkaloids is used in their extraction and further clean-up as shown in schem (10)[14].

The methanolic extract which gave negative test with Dragendorff's reagent was brought to 2N HCl and reduced with Zn dust with stirring overnight to convert Pyrrolizidine alkaloids N-oxides to the corresponding free bases[93, 139-141].

Excess Zn was removed by filtration. The aqueous acidic solution was washed with CH<sub>2</sub>Cl<sub>2</sub>, and made alkaline with NH<sub>4</sub>OH Using pH meter to make sure that pH in base range, After basification process, the solution was extracted with ethylene chloride, then dried using anhydrous sodium sulfate (Na<sub>2</sub>SO<sub>4</sub> 50 to 100 g/L organic layer) with stirring for three minutes using glass rod and allowed to stand for 5 minutes then filtered. Organic layer was then combined and concentrated under vacuum at 30 °C using rotary evaporator to obtain the

total alkaloids (tertiary Pyrrolizidine alkaloids and Pyrrolizidine alkaloids N-oxides) in form of tertiary Pyrrolizidine alkaloids[93, 139-142]. Alkaloid extracts were dissolved in methanol and analyzed by GC-MS.

#### Plant material



Extraction with Methanol to produce

MeOH Fraction



MeOH fraction dissolved in 0.5 M HCland remove undissolved material and reduced with Zn dust



Washed with dichloromethane to remove non-alkaloids such as fats, wax and other organic compounds



Basified the aqueous solution with (25%) ammonia



Extraction with dichloromethane to Produce total alkaloids

Scheme 10: Extraction scheme of Pyrrolizidine alkaloids from Echium plantagineum

#### 2.6. GC-MS analysis

Aglient 6890 chromatograph equipped with aglient mass spectrometric detector, with a direct capillary interface and fused silica capillary column HP-5MS (30 m  $\times$  0.32 mm  $\times$  0.25  $\mu$ m film thickness).

Samples were injected under the following conditions:

Helium was used as carrier gas at approximately 1.0 ml/min, pulsed splitless mode. The solvent delay was 3 min. and injection size of sample was 0.001 ml. The mass spectrometric detector was operated in electron impact ionization mode with energy of 70 ev. Scanning from m/z 50 to m/z 500. The ion source temperature was 230 °C. and the quadruple temperate was 150 °C. the electron multiplier voltage (EM voltage) was maintained 1050 v above auto tune. The GC temperature program was started at 70 °C then elevated to 300 °C at rate 6 °C/min.

### 2.6.1. Kovats Retention indices (RI)

Kovats retention index RI is a linear scale based upon thermodynamic properties and observation of chromatographic data trends that show a distinct advantage in predicting retention time. The system uses n-alkanes as an index against which retention times of other compounds can be measured. The retention index is information unique to a compound [143].

The temperture programed Kovats retention index RI is defined for given substance (A) as follow[144, 145]

$$RI_{A} = 100 \left( \frac{t_{rA} - t_{rn}}{t_{rN} - t_{rn}} + n \right)$$

trA is retention time of unknown substance (A).

trn is retention time of n-alkane elute before (A).

trN is retention time of n-alkane elute after (A).

n and N refer to the carbon numbers in n-alkane.

Unlike retention time, retention index of compounds has the following advantages [146, 147].

- (i) Its dependence on temperature is small and linear.
- (ii) It is independence of the column constants and of type of chromatographic apparatus.
- (iii)It provide chemical information about the chemical nature of the substances under examination.

#### 2.7. Reagents

- 1. Petroleum ether, Analytical grade, range of boiling 45-60 °C, Scharlau Chemie S.A. Barcelona, Spain.
- 2. Dichloromethane, Synthesis grade, range of boiling 39-40 °C. 99.5%. mgchemicals, Belgium. . CAS No. 75-09-2.
- 3. Ammonia solution. About 25% NH<sub>3</sub>. Chemically pure. Frutarom (UK) LTD. CAS No. 1336-21-6.
- 4. Ethanol, Synthesis grade, 99.9%. Scharlau Chemie S.A. Barcelona, Spain.
- 5. Methanol, analytical grade, boiling range 64-65.5 °C. 99.5%. BDH England.
- 6. Acetone, Analytical grade, Laboratory Chemicals, England.
- 7. Chloroform, pure. Lonover Laboratory Chemicals, England.
- 8. Anhydrous sodium sulphate, Laboratory reagent. India

#### 2.8. Instrumentation

- 1. Rotary Evaporator, RE 200, BIBBY, made in U.K. by BIBBY STERILINE LTD.
- 2. UV lamp, Herolab, made in Germany by GmbH laborgeate K W-254 nm.
- 3. JENWAY pH meter 3310.

## **CHAPTER THREE**

#### 3. RESULTS AND DISCUSSION

#### 3.1. Structural Elucidation of Isolated Compounds

Gas chromatography combined with mass spectrometry has been increasingly used over last two decades for the convenient analysis of alkaloidal extracts containing pyrrolizidine alkaloids [147-150]. The combination of retention indices (RI), molecular mass [M<sup>+</sup>], and mass fragmentation pattern provide sufficient information for unambiguous identification of most pyrrolizidine alkaloids[138, 147].

#### 3.2. GC-MS analysis of the alkaloidal extract of *Echium plantagineum* L

Gas chromatography combined with mass spectrometry has been increasingly used over last two decades for the convenient analysis of alkaloidal extracts containing pyrrolizidine alkaloids [147-150]. This method of choice for the analysis of crude alkaloidal fraction which contain about 10 individual components and enables the identification of even trace or stereoisomers. The combination of retention indices (RI), molecular mass [M<sup>+</sup>], and mass fragmentation pattern provide sufficient information for unambiguous identification of most pyrrolizidine alkaloids.

GC analysis of the alkaloidal extract of *Echium plantagineum* L revealed the presence of 12 alkaloids of pyrrolizidine type as illustrated in table (2). Figure (7) shows the total profile obtained by GC-MS of the alkaloidal extract of *Echium plantagineum* L.

Kovats retention indices of isolated pyrrolizidine alkaloids were calculated using cochromatographed standard n-alkanes (C17- C33). The retention index of data were calculated with the respect to the separately injected standard hydrocarbon illustrated in table (3) and figure (8).

Peak No	Alkaloid	Retention time (min)	Retention index
1	Ech-1	4.53	1787
2	Ech-2	4.7	1797
3	Ech-3	5.46	1843
4	Ech-4	10.75	2133
5	Ech-5	10.94	2145
6	Ech-6	13.01	2255
7	Ech-7	14.21	2315
8	Ech-8	14.38	2325
9	Ech-9	14.51	2533
10	Ech-10	14.58	2337
11	Ech-11	18.72	2560
12	Ech-12	30.24	3178

Table 2: GC data of the alkaloidal extract of Echium plantagineum L

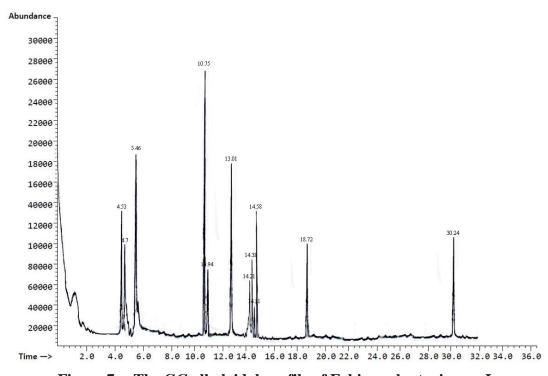


Figure 7: The GC alkaloidal profile of Echium plantagineum L

Peak No	n-Alkane	Retention time (min)
1	C-17	3.05
2	C-18	4.75
3	C-19	6.41
4	C-20	8.15
5	C-21	10.2
6	C-22	11.85
7	C-23	13.95
8	C-24	15.65
9	C-25	17.7
10	C-26	19.4
11	C-27	21.1
12	C-28	23.2
13	C-29	24.8
14	C-30	26.9
15	C-31	28.6
16	C-32	30.7
17	C-33	32.4

Table 3: GC data of the injected standard hydrocarbon

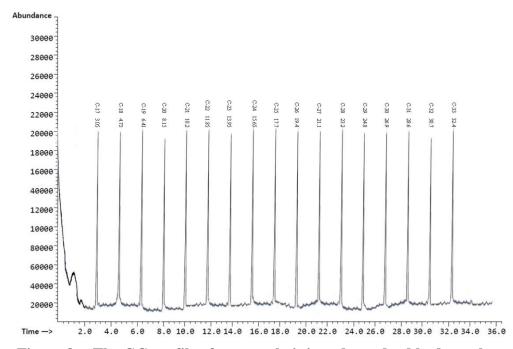


Figure 8: The GC profile of separately injected standard hydrocarbon

The table (4) shows the pyrrolizidine alkaloids that have been identified using GC-MS technique; these compounds has been used as a standards for elucidate and identify the separated compounds in these research.

Alkaloid	RI	Formula	<b>M</b> <sup>+</sup>	Characteristic ions(%)	ref
		03		237(2), 219 (3), 204(0.5), 191(1), 154(2), 138(5), 137(23),	
7-Angeloylretronecine	1787	$C_{13}H_{19}NO_3$	237	136(18), 124(23), 111(38),	[151]
		$C_{13}$		106(40), 94(20), 93(6), 83(11),	
				80(100), 55(22).	
				237(1), 219(0.5), 193(3),	
		$C_{13}H_{19}NO_3$		154(16), 138(32), 137(25),	
9-Angeloylretronecine	1797	$H_{19}$	237	136(10), 126(7), 120(2), 108(2),	[151]
		$C_{13}$		94(25), 93(100), 83(8), 80(10),	
				55(13).	
	1843	)3		193(5), 154(15), 138(20),	
9-Tigloylretronecine		C <sub>13</sub> H <sub>19</sub> NO <sub>3</sub>	237	137(26), 136(13), 126(7),	[151]
			13H <sub>1</sub>	119(5), 109(4), 94(23), 93(100),	[101]
		S		83(10), 80(12), 55(18).	
		)5		299(0.4), 156(9), 139(35),	
Intermedine	2133	<sub>5</sub> NC	299	138(100), 137(13), 136(13),	[47]
memediie	2133	C <sub>15</sub> H <sub>25</sub> NO <sub>5</sub>	2))	120(10), 95(15), 94(50), 93(80),	[ דיי ]
		ت		80(14), 67(9), 45(7), 43(18).	
				299(0.5), 254(1), 156(8),	
		05		139(31), 138 (100), 137(12),	
Lycopsamine	2145	C <sub>15</sub> H <sub>25</sub> NO <sub>5</sub>	299	136(12), 120(10), 108(4),	[100]
		$\mathbb{C}_{15}\mathbf{F}$		95(15), 94 (55), 93(84), 80(14),	
				67(10), 45(8), 43(20).	

Acetylintermedine	2255	C <sub>17</sub> H <sub>27</sub> NO <sub>6</sub>	341	341(5), 298 (4), 255 (16), 139 (20), 138(100), 137 (12),136 (12), 94 (30), 93 (71), 80 (10), 43 (21).	[47]
7-Angeloyl-9-(2,3- dihydroxybutyryl) retronecine	2315	$C_{17}H_{25}NO_6$	339	339(1), 239(5), 238(5), 237(5), 221(25), 220(99), 219(15), 141(20), 138(10), 137(11), 136 (100), 121(15), 120(83), 119(34), 106(10), 94(55), 93(95), 83(41), 80(20), 75(2), 57(10), 55(40), 45(10).	[151]
7-Tigloyl-9-(2,3-dihydroxybutyryl) retronecine	2325	C <sub>17</sub> H <sub>25</sub> NO <sub>6</sub>	339	239(5), 238(5), 237(8), 221(22), 220(90), 219 (20), 141(20), 138(10), 137(11), 136(100), 121(15), 120(80), 119(35), 106(10), 94(58), 93(90), 83(46), 80(20), 75(2), 57(9), 55(40), 45(10).	[151]
7-Angeloyl-9-(2,3- dihydroxybutyryl) heliotridine	2333	$\mathrm{C_{17}H_{25}NO_{6}}$	339	339(1), 324(1), 294(1), 239(6), 222(25), 221(25), 220(65), 219(8), 138 (20), 137(10), 136(81), 121(24), 120(100), 119(85), 106(15), 94(50), 93(85), 83(24), 80(18), 75(2), 57(10), 55(25), 45(10).	[152]

Uplandicine	2337	C <sub>17</sub> H <sub>27</sub> NO <sub>7</sub>	357	357(4), 342(5), 297(23), 281(4), 256(4), 207 (7), 206(52), 181(80), 180 (100), 179 (43), 136(75), 121(49), 120(85), 119 (50), 101(23), 94(55), 93(74), 80(28), 73(74), 59(23), 45(20), 44(69), 43(48).	[153]
Echimidine	2560	C <sub>20</sub> H <sub>31</sub> NO <sub>7</sub>	397	397(0.1), 382 (0.1), 352 (0.1), 297(2), 221(21), 220(100), 219(5), 138(5), 137(6), 136(48), 121(26), 120(75), 119(30), 106(5), 94(30), 93 (61), 83(39), 80(10), 59(10), 55(25), 43(18).	[151]
Echiumine	3178	C <sub>20</sub> H <sub>31</sub> NO <sub>7</sub>	381	381(0.4), 338(1), 337(1), 336(1), 281(2), 255(1), 238(9), 237(2), 221(35), 220(100), 141(16), 138(8), 136(47), 121(32), 120(69), 119(20), 106(7), 94(39), 93(50), 83(28), 80(17), 59(1), 55(37), 53(9).	[154]

Table 4: GC-MS data of the standard Pyrrolizidine alkaloids previously separated

### 3.3. Characterization of Ech-1 pyrrolizidine alkaloid

The mass spectrum of Ech-1 pyrrolizidine alkaloid figure (9) showed molecular ion peak at m/z 237 for  $C_{13}H_{19}NO_3$ . The mass spectra data table (5) showed significant ions at m/z 136, m/z 120, m/z 119, m/z 93, and 80 m/z. these fragments are characteristic to unsaturated necine base [22, 24, 138, 155], the intense ion at m/z 137 (M<sup>+</sup>-angelic acid) and m/z 106 are characteristic for C-7 monoester; which corresponding to the loss of ester and primary alcohol group respectively [138, 155] as shown in scheme (11), the ion at m/z 219 is due to the loss of water molecule (M<sup>+</sup>-H<sub>2</sub>O)

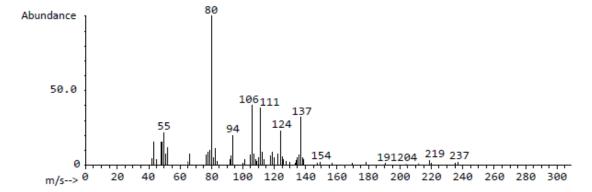
According to data given in literature [99, 151]The alkaloid Ech-1 could be unambiguously identified by its specific retention index and its mass fragmentation pattern as 7-angeloylretronecine

7-angeloylretronecine

m/z	Relative abundance (%)	Expected formula
237	2	$C_{13}H_{19}NO_3$
219	3	$C_{13}H_{17}NO_2$
154	2	$C_8H_{12}NO_2$
137	23	C <sub>8</sub> H <sub>11</sub> NO
124	23	$C_7H_{10}NO$
111	38	$C_6H_9NO$
106	40	C <sub>7</sub> H <sub>8</sub> N
94	20	$C_6H_9N$
83	11	C <sub>5</sub> H <sub>7</sub> O
80	100	$C_5H_6N$
55	22	C <sub>4</sub> H <sub>7</sub>

Table 5: the mass spectral data of Ech-1

Scheme 11: A fragmentation pathway for Ech-1 alkaloid



Mass spectrum of Ech-1 alkaloid

#### 3.4. Characterization of Ech-2 and Ech-3 pyrrolizidine alkaloids

The Ech-2 and Ech-3 alkaloids both exhibit approximately the same fragmentation pattern in mass spectrum figure (10 and 11) only small different occurred in relative intensities of some fragment ions tables (6).

Apparently both Ech-2 and Ech-3 alkaloids have a molecular mass of 237 and observe the characteristic peaks for monoesters of an unsaturated base, viz at m/z 80, 93, 136, 136 and 138[22, 24, 138, 151, 156]. An intense peak at m/z 138 indicates that the compounds are 9monoesters[22, 70-72] scheme (12). By comparison of retention indices and mass spectra to data given in literature[151], both alkaloids were tentatively identified as 9angelylretronecine for Ech-2, and 9-tigelylretronecine for Ech-3 alkaloids.

The tigloyl esters are delayed during GC over the angeloyl esters. This is due to trans configuration of the carbonyl group and methyl group on tigloylesters versus the cisconfiguration on the angeloyl esters[147, 157].

9-Tigeloylretronecine

71

	Relative al	Expected	
m/z	Ech-2	Ech-3	formula
237	1	1	C <sub>13</sub> H <sub>19</sub> NO <sub>3</sub>
219	0.5	0.4	C <sub>13</sub> H <sub>17</sub> NO <sub>2</sub>
138	32	20	C <sub>8</sub> H <sub>12</sub> NO
137	25	26	C <sub>8</sub> H <sub>11</sub> NO
136	10	13	C <sub>8</sub> H <sub>10</sub> NO
119	5	5	C <sub>8</sub> H <sub>9</sub> N
120	2	2	$C_8H_{10}N$
94	25	23	C <sub>6</sub> H <sub>8</sub> N
93	100	100	C <sub>6</sub> H <sub>7</sub> N
83	8	10	C <sub>5</sub> H <sub>7</sub> O
80	10	12	C <sub>5</sub> H <sub>6</sub> N

Table 6: the mass spectral data of Ech-2 and Ech-3

Scheme 12: A fragmentation pathway for Ech-2 and Ech-3 alkaloids

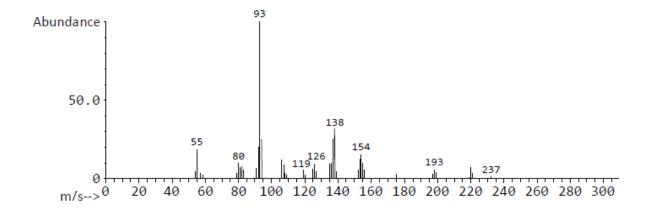


Figure 10: Mass spectrum of Ech-2 alkaloid

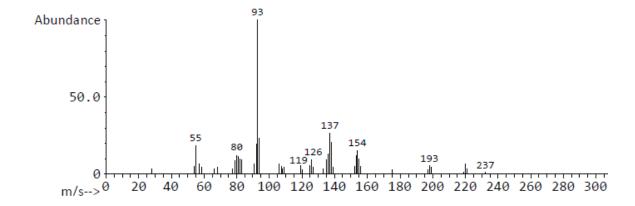


Figure 11: Mass spectrum of Ech-3 alkaloid

# 3.5. Characterization of Ech-4 and Ech-5 pyrrolizidine alkaloid

Ech-4 and Ech-5 alkaloids both show a molecular ion peak at m/z 299 figures (12) and (13), which corresponds to the molecular formula  $C_{15}H_{25}NO_5$  tables (7). The Pyrrolizidine alkaloidse peak m/z 138 is due to cleave of the weak allylic ester bond scheme (13) give strong evidence for the presence of free OH group at C-7[22, 70-72, 155].

significant ions m/z 136, m/z 94, m/z 93, and m/z 80 is characteristic to 1,2 unsaturated necine base[22, 24, 47, 99, 138, 155, 158].

Comparison of retention indices and mass fragmentations with those reported in literature [147, 159, 160] show clearly that Ech-4 alkaloid is Intermedine and Ech-5 alkaloid is lycopsamine.

/rz	Relative abu	indance (%)	Expected formula
m/z	Ech-4	Ech-5	Expected formula
299	0.4	0.5	C <sub>15</sub> H <sub>25</sub> NO <sub>5</sub>
139	35	31	C <sub>8</sub> H <sub>13</sub> NO
138	100	100	C <sub>8</sub> H <sub>12</sub> NO
137	13	12	C <sub>8</sub> H <sub>11</sub> NO
136	13	12	C <sub>8</sub> H <sub>10</sub> NO
120	10	10	$C_8H_{10}N$
94	50	55	C <sub>6</sub> H <sub>8</sub> N
93	80	84	C <sub>6</sub> H <sub>7</sub> N
80	14	14	C <sub>5</sub> H <sub>6</sub> N
45	7	8	C <sub>2</sub> H <sub>5</sub> O
43	43	20	C <sub>3</sub> H <sub>7</sub>

Table 7: the mass spectral data of Ech-4 and Ech-5

Scheme 13: A fragmentation pathway for Ech-4 and Ech-5 alkaloids

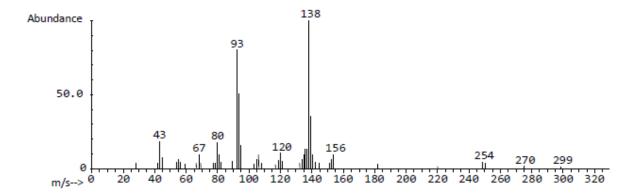


Figure 12: Mass spectrum of Ech-4 alkaloid

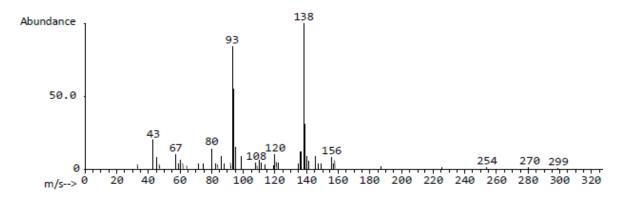


Figure 13: Mass spectrum of Ech-5 alkaloid

# 3.6. of Ech-6 pyrrolizidine alkaloid

The mass spectrum figure (14) showed  $M^+$  at m/z 341, this parent ion is corresponding to the following molecular formula  $C_{17}H_{27}NO_6$  table (8).

The fragment ions m/z 136, m/z 94, m/z 93, and m/z 80 is characteristic to 1,2 unsaturated necine base [22, 24, 47, 99, 138, 155, 158].

The peak at m/z 138 is formed through C-9 cleavage scheme (14), this peak is characteristic for C9-monoesters [22, 70-72, 155].

A base peak at m/z 138 due to cleavage of the weak allylic ester bond, providing strong evidence for the presence of a free OH at C-7[100, 161]

Comparison of retention index and mass fragmentations with those reported in literature [47, 99] show that the Ech-6 alkaloid is acetylderivative of Intermedineor acetylderivative of lycopsamine. However, it needs to be established by further technique whether the acetyl group is in the 3' or 2' position.

3'-acetylintermedine

3'-acetylycopsamine

2'-acetylintermedine

2'-acetylycopsamine

m/z	Relative abundance (%)	Expected formula
341	5	C <sub>17</sub> H <sub>27</sub> NO <sub>6</sub>
138	100	C <sub>8</sub> H <sub>12</sub> NO
137	12	C <sub>8</sub> H <sub>11</sub> NO
136	12	C <sub>8</sub> H <sub>10</sub> NO
94	30	C <sub>6</sub> H <sub>8</sub> N
93	71	C <sub>6</sub> H <sub>7</sub> N
80	10	C <sub>5</sub> H <sub>6</sub> N
43	21	C <sub>2</sub> H <sub>3</sub> O

Table 8: the mass spectral data of Ech-6

HO OH HO OH HO OH HO OH HO OH MIX. 255 
$$C_{2}H_{3}O^{+}$$
  $C_{17}H_{27}NO_{6}^{++}$   $C_{17}H_{27}NO_{6}^{++}$   $C_{17}H_{27}NO_{6}^{++}$   $C_{2}H_{3}O^{+}$   $C_{2}H_{3}O^{+}$   $C_{2}H_{3}O^{+}$   $C_{2}H_{3}O^{+}$   $C_{2}H_{3}NO^{++}$   $C_{3}H_{13}NO^{++}$   $C_{3}H_{13}NO^{++}$   $C_{4}H_{13}NO^{++}$   $C_{5}H_{6}N^{+}$   $C_{5}H_{$ 

Scheme 14: A fragmentation pathway for Ech-6 alkaloid

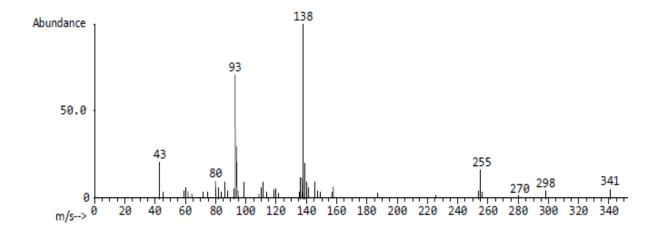


Figure 14: Mass spectrum of Ech-6 alkaloid

#### 3.7. Characterization of Ech-7, Ech-8 and Ech-9 pyrrolizidine alkaloid

Ech-7,Ech-8, and Ech-9 alkaloids all have molecular ion peak at m/z 339 which corresponding to the formula ( $C_{17}H_{25}NO_6$ ) tables (9) respectively. And they show the fragmentation pattern of un saturated diester pyrrolizidine alkaloid which are m/z136, m/z 120, m/z 119, m/z 93, and m/z 80[22, 24, 47, 99, 138, 155, 158].

The strong peak at m/z  $220(M^+-C_4H_7O_4)$  is due to the cleavage of the weak allylic bond at C-9.and the ion peak at m/z  $239(M^+$ -angelic acid) is due to the loss of acid attached to C-7 scheme (15).

The fragment at m/z 57 ( $C_3H_5O$ ) is probably derived from m/z 75 ( $C_3H_7O_2$ ) through loss one molecule of water.

Ech-7, Ech-8, and Ech-9 alkaloids show similar fragmentation pattern figures (15), (16), and (17) respectively but they differ in the retention indices and in the relative intensities of some fragment ion.

Ech-7 alkaloid was tentatively identified as 7-Angeloyl-9-(2,3-dihydroxybutyryl)retronecine according to literature data [95, 99, 147, 152], and alkaloids Ech-8 and Ech-9 were assumed to be isomers of Ech-7.

m/z	Relative	e abunda	nce (%)	Expected
III/Z	Ech-7	Ech-8	Ech-9	formula
339	1	1	1	C <sub>17</sub> H <sub>25</sub> NO <sub>6</sub>
239	5	5	6	C <sub>12</sub> H <sub>17</sub> NO <sub>4</sub>
221	25	25	25	$C_{13}H_{19}NO_2$
220	99	90	65	$C_{13}H_{18}NO_2$
136	100	100	81	C <sub>8</sub> H <sub>10</sub> NO
120	83	80	100	$C_8H_{10}N$
119	34	35	85	C <sub>8</sub> H <sub>9</sub> N
106	10	10	15	C <sub>7</sub> H <sub>8</sub> N
94	55	58	50	C <sub>6</sub> H <sub>9</sub> N
93	95	90	85	C <sub>6</sub> H <sub>7</sub> N
83	41	46	24	C <sub>5</sub> H <sub>7</sub> O
55	40	40	25	$C_4H_7$

Table 9: the mass spectral data of Ech-7, Ech-8, and Ech-9

Scheme 15: A fragmentation pathway for Ech-7, Ech-8 and Ech-9 alkaloids

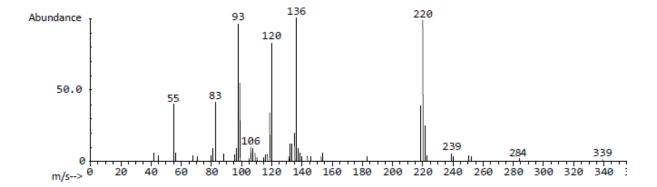


Figure 15: Mass spectrum of Ech-7 alkaloid

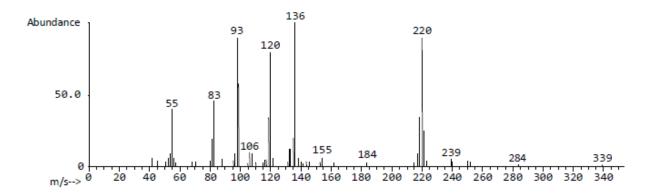


Figure 16: Mass spectrum of Ech-8 alkaloid

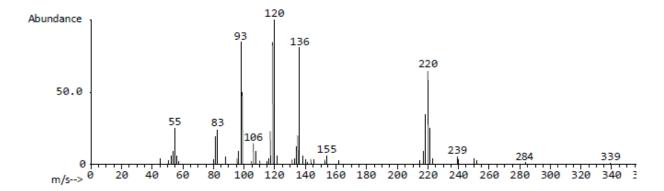


Figure 17: Mass spectrum of Ech-9 alkaloid

### 3.8. Characterization of Ech-10 pyrrolizidine alkaloid

Figure (18) shows M+ at m/z 357 (corresponding to molecular formula  $C_{17}H_{27}NO_7$ ) table (10). The ion series at m/z 136, 120, 119, 93, and 80 are characteristic of 1.2-unsaturated diester pyrrolizidine alkaloids[22, 24, 47, 99, 138, 155, 158], and the base peak at m/z 180 (M- $C_7H_{13}O_5$ ) provides strong evidence for the presence of C-7 acetoxy group[154, 155]. The low intensity peak at m/z 297 is due to lose of acid attached at C-7 (M- $C_2H_3O_2$ ) scheme (16).

By direct comparison of retention indice of Ech-10 alkaloid (RI) with literature data[152], The acid attached at C-9 has formula of ( $C_7H_{13}O_5$ ) can be identified as 5'-OH (+)-Trach (Echimidinyl), hence the alkaloid Ech-10 with RI 2337was identified as 7-acetyl-9-echimidinylretronecine (uplandicine).

m/z	Relative abundance (%)	Expected formula
357	4	C <sub>17</sub> H <sub>27</sub> NO <sub>7</sub>
297	23	$C_{15}H_{23}NO_5$
181	80	$C_{10}H_{15}NO_2$
180	100	$C_{10}H_{14}NO_2$
179	43	$C_{10}H_{13}NO_2$
136	75	C <sub>8</sub> H <sub>10</sub> NO
121	49	C <sub>8</sub> H <sub>11</sub> N
120	85	$C_8H_{10}N$
119	50	C <sub>8</sub> H <sub>9</sub> N
93	74	C <sub>6</sub> H <sub>7</sub> N
80	28	C <sub>5</sub> H <sub>6</sub> N
43	20	C <sub>2</sub> H <sub>3</sub> O

Table 10: the mass spectral data of Ech-10

Scheme 16: A fragmentation pathway for Ech-10 alkaloid

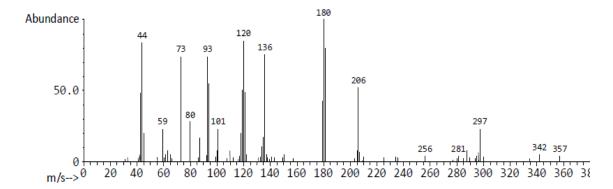


Figure 18: Mass spectrum of Ech-10 alkaloid

### 3.9. Characterization of Ech-11 pyrrolizidine alkaloid

The MS spectrum of Ech-7 figure (19) showed Molecular ion peak at m/z 397; where the relative abundance is 0.1 % for  $C_{20}H_{31}NO_7$ . The mass spectral data table (11) Showed significant ions at m/z 136, m/z 120, m/z 119, m/z 93, and m/z 80. These fragments are characteristic to 1,2-unsaturated necine base[22, 24, 138, 151, 156], the base peak ion at m/z 220 is formed through C-9 cleavage of diesterpyrrolizidine alkaloid bearing angeloxy group.

This result was confirmed by the presence of the fragment ion at m/z 297 (M<sup>+</sup>-angelic acid) as shown in scheme (17) [138, 162]

Echimidine

m/z	Relative abundance (%)	Expected formula
397	0.1	C <sub>20</sub> H <sub>31</sub> NO <sub>7</sub>
297	2	C <sub>15</sub> H <sub>23</sub> NO <sub>5</sub>
221	21	C <sub>13</sub> H <sub>19</sub> NO <sub>2</sub>
220	100	C <sub>13</sub> H <sub>18</sub> NO <sub>2</sub>
136	48	$C_8H_{10}N_O$
120	75	$C_8H_{10}N$
119	30	C <sub>8</sub> H <sub>9</sub> N
106	5	C <sub>6</sub> H <sub>7</sub> N
94	30	C <sub>6</sub> H <sub>8</sub> N
93	61	C <sub>6</sub> H <sub>7</sub> N
83	39	C <sub>5</sub> H <sub>7</sub> O
80	10	C <sub>5</sub> H <sub>6</sub> N
55	25	C <sub>4</sub> H <sub>7</sub>

Table 11: the mass spectral data of Ech-11

Scheme 17: A fragmentation pathway for Ech-11 alkaloid

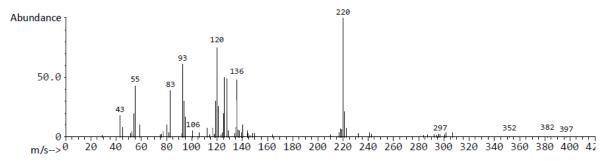


Figure 19: Mass spectrum of Ech-11 alkaloid

### 3.10. Characterization of Ech-12 pyrrolizidine alkaloid

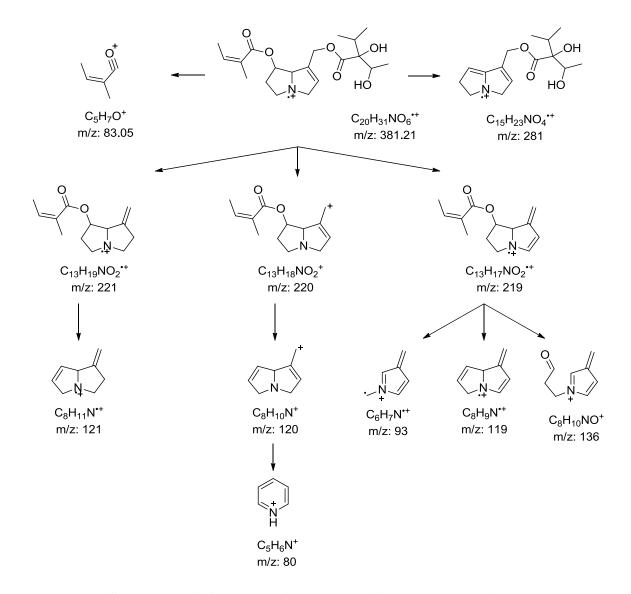
Ech-12 alkaloids have a molecular mass of 381 figure (20), and exhibit the characteristic peaks of an unsaturated Nicene base (m/z 80, 93, 119, 120, 121 and 136) [22, 24, 138, 151, 156]. Peaks at m/z 83 and 220 indicate the presence of an angelyl type group, while a peak at m/z 281 indicates a(±)- Trachelanthyl group[84]. The peak at m/z 220 is significantly more intense than that at m/z 281, suggesting that the(±)-Trachelanthyl group is attached at C-9 and the angelyl at C-7 [84]. This group of diesters is thus isomeric, with angelyl or tiglyl groups at C-7 and (±)-Trachelanthyl or (±)-Viridifloryl groups at C-9.

The retention index of Ech-12 is 3179. Comparison this value and mass spectra with those of standard alkaloids identified by GC-MS and other techniques in literature [154, 163] indicate that the Ech-12 combound is Echiumine which is a diester alkaloids and of unsaturated necine base.

Echiumine

m/z	Relative abundance (%)	Expected formula
397	0.4	C <sub>20</sub> H <sub>31</sub> NO <sub>7</sub>
221	35	C <sub>13</sub> H <sub>19</sub> NO <sub>2</sub>
220	100	$C_{13}H_{18}NO_2$
136	47	C <sub>8</sub> H <sub>10</sub> NO
120	69	$C_8H_{10}N$
119	20	C <sub>8</sub> H <sub>9</sub> N
106	7	C <sub>7</sub> H <sub>8</sub> N
94	39	C <sub>6</sub> H <sub>9</sub> N
93	50	C <sub>6</sub> H <sub>7</sub> N
83	28	C <sub>5</sub> H <sub>7</sub> O
80	17	C <sub>5</sub> H <sub>6</sub> N
55	37	C <sub>4</sub> H <sub>7</sub>

Table 12: the mass spectral data of Ech-12



Scheme 18: A fragmentation pathway for Ech-12 alkaloid

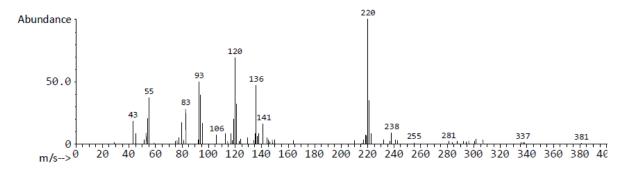


Figure 20: Mass spectrum of Ech-12 alkaloid

#### REFERENCES

- 1. Spainhour, C.B., *Natural Products*, in *Drug Discovery Handbook*. 2005, John Wiley & Sons, Inc. p. 11-72.
- 2. The classes of natural product and their isolation, in Natural Products: The Secondary Metabolites, J.R. Hanson, Editor. 2003, The Royal Society of Chemistry. p. 1-34.
- 3. McMurry, J.E., Secondary Metabolites: An Introduction to Natural Products Chemistry, in Organic Chemistry: With Biological Applications 2014 Cengage Learning.
- 4. Carocho, M. and I.C.F.R. Ferreira, *The Role of Phenolic Compounds in the Fight against Cancer*. Anti-Cancer Agents in Medicinal Chemistry, 2013,(13): p. 1236-1258.
- 5. Martins, D. and C.V. Nunez, *Secondary Metabolites from Rubiaceae Species*. Molecules, 2015. **20**: p. 13422-13495.
- 6. Petterson, D.S., D.J. Harris, and D.G. Allen, *chapter 7 Alkaloids*, in *Toxic Substances in Crop Plants*, J.P.F. D'Mello and C.M.D.H. Duffus, Editors. 1991, Woodhead Publishing. p. 148-179.
- 7. Natural Products (Secondary Metabolites), in Biochemistry & Molecular Biology of Plants, B. Buchanan, W. Gruissem, and R. Jones, Editors. 2000, John Wiley and Sons Ltd: Hoboken, United States. p. 1250-1318.
- 8. Kar, A., *Alkaloids*, in *Pharmacognosy and Pharmacobiotechnology*. 2003, New Age International (P) Ltd. p. 372-546.
- 9. Ng, Y.P., T.C. Or, and N.Y. Ip, *Plant alkaloids as drug leads for Alzheimer's disease*. Neurochem Int, 2015. **89**: p. 260-70.
- 10. Zimmermann, T.J., Design, Synthesis and Evaluation of Natural Product-Based Compound Collections, in Department of chemistry. 2012, Technischen Universität Dortmund. p. 220.
- 11. Dewick, P.M., *ALKALOIDS*, in *Medicinal Natural Products A Biosynthetic Approach*. 2002 John Wiley & Sons Ltd.
- 12. Kittakoop, P., C. Mahidol, and S. Ruchirawat, *Alkaloids as important scaffolds in therapeutic drugs for the treatments of cancer, tuberculosis, and smoking cessation.* Curr Top Med Chem, 2014. **14**(2): p. 239-52.
- 13. Qiu, S., et al., *Natural alkaloids: basic aspects, biological roles, and future perspectives.* Chin J Nat Med, 2014. **12**(6): p. 401-6.
- 14. Ibrahim, E.S., *Isolation And Characterization Of Pyrrolizidine Alkaloids From Echium Glomeratum Poir (Boraginaceae)*. 2007, Jordan University of Science and Technology.
- 15. Evans, W.C. and G.E. Trease, *Alkaloids*, in *Trease and Evans pharmacognosy*. 2002, W.B. Saunders. p. 333-393.
- 16. Kumar, S., *Alkaloidal Drugs A Review*. Asian Journal of Pharmaceutical Science & Technology, 2014 **4**(3): p. 107-119.

- 17. Djilani, A., et al., *New extraction technique for alkaloids*. Journal of the Brazilian Chemical Society, 2006. **17**: p. 518-520.
- 18. Raaman, N., *Qualititative phytochemical sceening* in *Phytochemical Techniques*. 2006, New India Publishing. p. 19-24.
- 19. Kakhia, T.I., *Alkaloids and Alkaloids Plants*, Adana University.
- 20. Alagarsamy, V., *Alkaloids*, in *Pharmaceutical Chemistry of Natural Products* 2012, Elsevier India. p. 149-176.
- 21. Aniszewski and Tadeusz, *Alkaloids secrets of life*. 2007: Amsterdam ; Oxford : Elsevier.
- 22. Grue, M.R., A Study Of The Alkaloid Content of the Senecio Speciosus / Macrocephalus Complex. 1991, Rhodes University.
- 23. Roeder, E., et al., *Pyrrolizidine alkaloids of Heliotropium spathulatum*. Phytochemistry, 1991. **30**(5): p. 1703-1706.
- 24. Mattocks, A.R., *Chemistry and Toxicology of Pyrrolizidine Alkaloids*. 1986: Academic Press Inc. (London) Ltd.
- 25. Cheng and Dandan, *Pyrrolizidine alkaloid variation in Jacobaea hybrids : influence on resistance against generalist and specialist insect herbivores.* Division Plant Ecology and Phytochemistry, 2012.
- 26. Fu, P.P., et al., *Pyrrolizidine Alkaloids—Genotoxicity, Metabolism Enzymes, Metabolic Activation, and Mechanisms*. Drug Metabolism Reviews, 2004. **36**(1): p. 1-55.
- 27. Asres, K., F. Sporer, and M. Wink, *Patterns of pyrrolizidine alkaloids in 12 Ethiopian Crotalaria species*. Biochemical Systematics and Ecology, 2004. **32**(10): p. 915-930.
- 28. Liddell and J. Richard, *Synthetic studies of swazinecic acid dilactone*, in *Faculty of Science, Chemistry*. 1989, Rhodes University: Rhodes p. 290.
- 29. Marquina, G., et al., *Antimicrobial activity of pyrrolizidine alkaloids from Heliotropium bursiferum Wr ex Grisebach.* Pharmazie, 1989. **44**(12): p. 870-1.
- 30. Van Wyk, B.-E. and G.H. Verdoorn, *A chemotaxonomic survey of major alkaloids in Lotononis and Buchenroedera*. Biochemical Systematics and Ecology, 1989. **17**(5): p. 385-389.
- 31. Borstel, K.V., L. Witte, and T. Hartmann, *Pyrrolizidine alkaloid patterns in populations of Senecio vulgaris, S. vernalis and their hybrids.* Phytochemistry, 1989. **28**(6): p. 1635-1638.
- 32. Meinwald, J., Alkaloids and isoprenoids as defensive and signalling agents among insects 1990. **62**(7): p. 1325-1328.
- 33. Nahrstedt, A., *The Significance of Secondary Metabolites for Interactions between Plants and Insects.* Planta Med, 1989. **55**(04): p. 333-338.
- 34. L'Empereur, K.M., et al., *Pyrrolizidine Alkaloids from Hackelia californica and Gnophaela latipennis, an H. californica-Hosted Arctiid Moth.* Journal of Natural Products, 1989. **52**(2): p. 360-366.
- 35. Schneider, M.J. and F.R. Stermitz, *Uptake of host plant alkaloids by root parasitic Pedicularis species*. Phytochemistry, 1990. **29**(6): p. 1811-1814.

- 36. TH, J., et al., Chemotaxonomic implications of the venom chemistry of someMonomorium "antarcticum" populations. Journal of Chemical Ecology, 1988 14(12): p. 2197-212.
- 37. Liddell and J. Richard, *Synthetic studies of Swazinecic Acid Dilactone*. 1988, Rhodes University.
- 38. GRUE, M.R., A Study of the Alkaloid Content of the Senecio Speciosus / Macrocephalus Complex. 1991, Rhodes University.
- 39. ROEDER, E., Medicinal plants in Europe containing pyrrolizidine alkaloids. Pharmazie, 1995. **50**(2): p. 83-98.
- 40. Acamovic, T., C.S. Stewart, and T.W. Pennycott, *Experiences with the Quantitative Trace Analysis of Pyrrolizidine Alkaloids using GCMS and LCMS*, in *Poisonous Plants and Related Toxins*, S.M.C. K.Beales and J.A. Edgar, Editors. 2004, Wallingford, Oxon, UK Cambridge, MA, USA: CABI Pub. p. 453-470.
- 41. Hartmann, T. and G. Toppel, Senecionine n-oxide, the primary product of pyrrolizidine alkaloid biosynthesis in root cultures of Senecio vulgaris. Phytochemistry, 1987. **26**(6): p. 1639-1643.
- 42. Boppre, M., *Lepidoptera and Pyrrolizidine Alkaloids Exemplification of Complexity In Chemical Ecology.* journal of chemical ecology, 1990. **16**(1).
- 43. Zhang, F., et al., *Quantitative analysis of total retronecine esters-type pyrrolizidine alkaloids in plant by high performance liquid chromatography*. Analytica Chimica Acta, 2007. **605**(1): p. 94-101.
- 44. Roeder, E., *Analysis of Pyrrolizidine Alkaloids*. Current Organic Chemistry, 1999. **3**: p. 557-576.
- 45. Dreger, M., et al., *Pyrrolizidine alkaloids chemistry, biosynthesis, pathway, toxicity, safety and perspectives of medicinal usage.* Herba Polonica, 2009. **55** (4): p. 127-147.
- 46. Bull, L.B., C.C.J. Culvenor, and A.T. Dick, *The pyrrolizidine alkaloids, North Holland.* 1968, Amsterdam.
- 47. El-Shazly, A., et al., *Pyrrolizidine alkaloids in members of the Boraginaceae from Sinai (Egypt)*. Biochemical Systematics and Ecology, 1998. **26**(6): p. 619-636.
- 48. El-Dahmy, S. and A. Adel Ghani, *Alkaloids of Arnebia decumbens Vent.* Az. J. Pharm. Sci, 1995. **15**: p. 24-34.
- 49. Roeder, E. and B. Rengel, *Pyrrolizidine alkaloids from Lithospermum erythrorhizon*. Phytochemistry, 1990. **29**(2): p. 690-693.
- 50. Röder, E., H. Wiedenfeld, and A. Pfitzer, *Doriasenine, a pyrrolizidine alkaloid from Senecio doria*. Phytochemistry, 1988. **27**(12): p. 4000-4001.
- 51. Mothes, K. and H.R. Schutte, *Biosynthese der Alkaloide*. (*Mit Beitragen von H. Bohm u. a.*) *Hrsg. von K. Mothes und H.R. Schutte*. 1969, Berlin, Deutscher Verlag der Wissenschaften VEB, 1969.
- 52. Böttcher, F., R.-D. Adolph, and T. Hartmann, *Homospermidine synthase, the first pathway-specific enzyme in pyrrolizidine alkaloid biosynthesis*. Phytochemistry, 1993. **32**(3): p. 679-689.

- 53. Robins, D.J., *A biogenetically patterned synthesis of the pyrrolizidine alkaloid trachelanthamidine*. Journal of the Chemical Society, Chemical Communications, 1982(22): p. 1289-1290.
- 54. Kunec, E.K. and D.J. Robins, *Application of 2H n.m.r. spectroscopy to study the incorporation of enantiomeric [2-2H]-labelled putrescines into the pyrrolizidine alkaloid retrorsine*. Journal of the Chemical Society, Perkin Transactions 1, 1987(0): p. 1089-1093.
- 55. Kunec, E.K. and D.J. Robins, *Pyrrolizidine alkaloid biosynthesis. Synthesis of 3H-labelled trachelanthamidine and isoretronecanol and their incorporation into three pyrrolizidine bases (necines)*. Journal of the Chemical Society, Perkin Transactions 1, 1989(8): p. 1437-1441.
- 56. Denholm, A.A., H.A. Kelly, and D.J. Robins, *Pyrrolizidine alkaloid biosynthesis*. *Synthesis of N-([4-14C]-4-aminobutyl)-1,2-didehydropyrrolidinium and its incorporation into different pyrrolizidine bases (necines)*. Journal of the Chemical Society, Perkin Transactions 1, 1991(8): p. 2003-2007.
- 57. Roeder, E., *Medicinal plants in Europe containing pyrrolizidine alkaloids* Pharmazie 1995. **50** p. 83-98.
- 58. Molyneux, R.J. and J.N. Roitman, *Specific detection of pyrrolizidine alkaloids on thin-layer chromatograms*. Journal of Chromatography A, 1980. **195**(3): p. 412-415.
- 59. Larson, K.M., M.R. Roby, and F.R. Stermitz, *Unsaturated Pyrrolizidines from Borage (Borago officinalis)*, a Common Garden Herb. Journal of Natural Products, 1984. **47**(4): p. 747-748.
- 60. Dodson, C.D. and F.R. Stermitz, *Pyrrolizidine Alkaloids from Borage (Borago officinalis) Seeds and Flowers*. Journal of Natural Products, 1986. **49**(4): p. 727-728.
- 61. Liddell, J.R., *Pyrrolizidine Alkaloids*. Natural Product Reports, 1995. **12**: p. 413.
- 62. Mehrabani, M., et al., *Toxic Pyrrolizidine Alkaloids of Echium Amoenum*. Daru, 2006. **14**(3): p. 122-127.
- 63. Domínguez, D.M., et al., *Pyrrolizidine alkaloids from Canarian endemic plants and their biological effects.* Biochemical Systematics and Ecology, 2008. **36**(3): p. 153-166.
- 64. Rösemann, G., Analysis of pyrrolizidine alkaloids in Crotalaria species by HPLC-MS/MS in order to evaluate, in Department of Paraclinical Sciences. 2006, University of Pretoria. p. 105.
- 65. Zhang, F., et al., *Quantitative Analysis by HPLC-MS*<sup>2</sup> of the Pyrrolizidine Alkaloid Adonifoline in Senecio scandens. Phytochemical Analysis, 2008. **19**: p. 25–31.
- 66. Bicchi, C., et al., Cyclodextrin derivatives in the gas chromatographic separation of racemic mixtures of volatile compounds X. 2,3-Di-O-ethyl-6-O-tert.-butyldimethylsilyl-β- and -γ-cyclodextrins. Journal of Chromatography A, 1996. **742**(1–2): p. 161-173.
- 67. Caniato, R., et al., Capillary Gas Chromatography/Positive and Negative Ion Chemical Ionization Mass Spectrometry on Pyrrolizidine Alkaloids of Senecio inaequidens Using Ammonia and Hydroxyl Ions as the Reagent Species. Journal of Natural Products, 1989. **52**(1): p. 32-41.

- 68. Molyneux, R.J., et al., *Australine, a Novel Pyrrolizidine Alkaloid Glucosidase Inhibitor from Castanospermum australe*. Journal of Natural Products, 1988. **51**(6): p. 1198-1206.
- 69. Mohanraj, S., et al., *Helifoline, a pyrrolizidine alkaloid from Heliotropium ovalifolium.* Phytochemistry, 1981. **20**(8): p. 1991-1995.
- 70. Asibal, C.F., L.T. Gelbaum, and L.H. Zalkow, *Pyrrolizidine Alkaloids from Heliotropium rotundifolium*. Journal of Natural Products, 1989. **52**(4): p. 726-731.
- 71. Davicino, J.G., M.J. Pestchanker, and O.S. Giordano, *Pyrrolizidine alkaloids from Heliotropium curassavicum*. Phytochemistry, 1988. **27**(3): p. 960-962.
- 72. Edgar, J.A., et al., *Callimorphine: identification and synthesis of the cinnabar moth "metabolite"*. Tetrahedron Letters, 1980. **21**(14): p. 1383-1384.
- 73. Bredenkamp, M.W. and A. Wiechers, *NMR-SPI: A reliable method for determining the mode of ester attachment in pyrrolizidine alkaloids*. Tetrahedron Letters, 1987. **28**(32): p. 3729-3732.
- 74. Malik, A. and K. Rahman, *Stereostructure of Subulacine-N-oxide: A New Pyrrolizidine Alkaloid from Heliotropium subulatum.* 1988. **27**(3): p. 707-711.
- 75. Smith, L., et al., *Crotaleschenine, an Alkaloid of <I>Crotalaria leschenaultii</I>*. Australian Journal of Chemistry, 1988. **41**(4): p. 429-436.
- 76. Yamada, K., et al., *Petasinine and petasinoside, two minor alkaloids possessing a new necine isolated from petasites japonicus maxim.* Tetrahedron Letters, 1978. **19**(46): p. 4543-4546.
- 77. Pestchanker, M.J., M.S. Ascheri, and O.S. Giordano, *Pyrrolizidine Alkaloids from Senecio subulatus and S. glandulosus*. Planta Med, 1985. **51**(02): p. 165-167.
- 78. Röder, E., H. Wiedenfeld, and U. Pastewka, *Pyrrolizidinalkaloide aus Senecio vernalis*. Planta Med, 1979. **37**(10): p. 131-136.
- 79. Asada, Y., T. Furuya, and . New pyrrolizidine alkaloids from LJgulariadentata. Chern. Pharm. Bull, 1984. **32**(2): p. 475
- 80. Birecka, H., et al., *Pyrrolizidine alkaloids of Heliotropium from Mexico and adjacent U.S.A.* Phytochemistry, 1980. **19**(3): p. 421-426.
- 81. Cava, M.P., et al., *Alkaloids of Cacalia floridana*. The Journal of Organic Chemistry, 1968. **33**(9): p. 3570-3573.
- 82. Resch, J.F., et al., *Biologically Active Pyrrolizidine Alkaloids From the True Forget-Me-Not, Myosotis scorpioides.* Journal of Natural Products, 1982. **45**(3): p. 358-362.
- 83. Hirschmann, G.S., S. Banerjee, and J. Jakupovic, *A new type of pyrrolizidine alkaloid from Senecio grisebachii Baker*. Revista latinoamericana de química, 1985. **16**(2): p. 109-110.
- 84. Roitman, J., *The pyrrolizidine alkaloids of Senecio triangularis*. Australian Journal of Chemistry, 1983. **36**(6): p. 1203-1213.
- 85. FAO, WHO, and CODEX (2011) Discussion Paper on Pyrrolizidine Alkaloids.
- 86. Roeder, E., H. Wiedenfeld, and R. Schraut, *Pyrrolizidine alkaloids from Alkanna tinctoria*. Phytochemistry, 1984. **23**(9): p. 2125-2126.
- 87. Pedersen, E., *Pyrrolizidine alkaloids in Danish species of the family Boraginaceae*. Arch Pharm Chem Sci Ed, 1975. **3**: p. 55-64.

- 88. BrochDue, A.I. Aasen, and A. J., *Alkaloids of Anchusa officinalis L. Identification of the pyrrolizidine alkaloid lycopsamine*. Acta Chem Scand, 1980 **34**(1): p. 75-77.
- 89. Hendriks, H., A.P. Bruins, and H.J. Huizing, *Detection of Curassavine and some related pyrrolizidine alkaloids in an anchusa officinalis strain by means of positive ion and negative ion chemical ionization GC/MS*. Biological Mass Spectrometry, 1988. **17**(2): p. 129-132.
- 90. Okusa, P.N., et al., *Absence of pyrrolizidine alkaloids in Cordia gilletii de wild (boraginaceae)*. Biochemical Systematics and Ecology, 2012. **41**(0): p. 1-2.
- 91. EL-shazly, A., et al., *Pyrrolizidine alkaloids of cynoglossum officinale and cynoglossum amabile (family Boraginaceae)*. Biochemical Systematics and Ecology, 1996. **24**(5): p. 415-421.
- 92. Pedersen, E., *Minor pyrrolizidine alkaloids from Cynoglossum officinale L.* Dan Tidsskr Farm, 1970. **44**(7): p. 287-91.
- 93. Alali, F.Q., et al., *Pyrrolizidine alkaloids from Echium glomeratum (Boraginaceae)*. Phytochemistry, 2008. **69**(12): p. 2341-2346.
- 94. EI-Shazlya, A., A.T. M. Abdel-Alla, and M. Winkb, *Pyrrolizidine Alkaloids from Echium rauwolfii and Echium horridum (Boraginaceae)*. 1999.
- 95. Roeder, E., K. Liu, and T. Bourauel, *Pyrrolizidine alkaloids from Echium pininana*. Phytochemistry, 1991. **30**(9): p. 3107-3110.
- 96. EI-Shazly, A., et al., *Pyrrolizidine alkaloids from Echium setosum and Echium vulgare*. J. Nat. Prod, 1996. **59**: p. 310-313.
- 97. Boppre, M., S.M. Colegate, and J.A. Edgar, *Pyrrolizidine Alkaloids of Echium vulgare Honey Found in Pure Pollen.* J. Agric. Food Chem, 2005. **53**: p. 594-600.
- 98. Sarg, T., et al., *Pyrrolizidine alkaloids from Echium angustifolium. Fitoterapia.* 1992. **63**: p. 466-468.
- 99. El-Shazly, A. and M. Wink, *Diversity of Pyrrolizidine Alkaloids in the Boraginaceae Structures, Distribution, and Biological Properties.* Diversity, 2014. **6**(2): p. 188.
- 100. El-Shazly, A., et al., *Pyrrolizidine and tetrahydroisoquinoline alkaloids from Echium humile*. Phytochemistry, 1996. **42**(1): p. 225-230.
- 101. Carvalho, J.C.B., et al., *Pyrrolizidine alkaloids in two endemic capeverdian Echium species*. Biochem. Syst. Ecol., 2013. **50**: p. 1-6.
- 102. Culvenor, C.C.J., J.A. Edgar, and L.W. Smith, *Pyrrolizidine alkaloids in Honey from Echium plantagineum L.* J. Agric. Food Chem, 1981. **29**: p. 958-960.
- 103. Wassel, G., et al., *Toxic pyrrolizidine alkaloids of certain Boraginaceae*. Acta Pharm. Suec, 1987. **24**: p. 199-204.
- 104. Roeder, E., H. Wiedenfeld, and K.J. Kaus, *Das Pyrrolizidinalkaloid Intermedin aus Cerinthe minor L.* Sci. Pharm, 1990. **58**: p. 9-13.
- 105. Akramov, S.T., et al., *Alkaloids of Senecio jacobaea, Heliotropium acutiflorum and H. transoxanum.* Khim. Prir. Soedin, 1966. **4**: p. 258.
- 106. Birecka, H., M.W. Frohlich, and L.M. Glickman, *Free and esterified necines in Heliotropium species from mexico and texas.* 1983. **22**(5): p. 1167-1171.
- 107. Zalkow, L.H., et al., *Pyrrolizidine alkaloid from middle eastern plans*. J. Nat. Prod, 1979. **42**: p. 603-614.

- 108. Catalfamo, J.L., W.B. Martin, and H. Birecka, *Accumulation of alkaloids and their necines in Heliotropium curassavicum*, H. spathulatum and H. indicum. Phytochemistry, 1982. **21**: p. 2669-2675.
- 109. Yassa, N.H., et al., *pyrrolizidine alkaloids of Heliotropium europaeum L. population Garmsar*. J. Sci. Islamic Repub. Iran, 1999. **10**: p. 39-42.
- 110. Tosun, F. and U. Tamer, *Determination of pyrrolizidine alkaloids in the seeds of Heliotropium europaeum by GC-MS*. J. Fac. Pharm. Ankara, 2004. **33**: p. 7-9.
- 111. Souza, J.S.N., et al., *Pyrrolizidine alkaloids from Heliotropium indicum*. J. Braz. Chem. Soc, 2005. **16**: p. 1410-1414.
- 112. Singh, J.P., et al., *Alkaloids of Heliotropium indicum*. J. Ind. Chem. Soc, 2005. **82**: p. 175-176.
- 113. Constantinidis, T., C. Harvala, and A.L. Skaltsounis, *Pyrrolizidine N-oxide alkaloids of Heliotropium hirsutissimum. Phytochemistry.* 1993. **32**: p. 1335-1337.
- 114. Dash, G.K. and M.S. Abdullah, *A review on Heliotropium indicum L. (Boraginaceae)* Int. J. Pharm. Sci. Res, 2013. **4**: p. 1253-1258.
- 115. Ravi, S., A.J. Lakshmanan, and W. Herz, *Isolycopsamine*, a pyrrolizidine alkaloid from Heliotropium keralense. Phytochemistry, 1990. **29**: p. 361-364.
- 116. Reina, M., et al., *Pyrrolizidine alkaloids from Heliotropium megalanthum*. J. Nat. Prod, 1998. **61**: p. 1418-1420.
- 117. Kiyamitdinova, F., S.T. Akramov, and S.Y. Yunusov, *Alkaloids from the family Boraginaceae*. Khim. Prir. Soedin, 1967. **3**: p. 411-412.
- 118. Mohanraj, S., et al., *Helifoline, a pyrrolizidine alkaloid from Heliotropium ovalifolium.* Phytochemistry 1981. **20**: p. 1991-1995.
- 119. Zalkow, L.H., L. Gelbaum, and E. Keinan, *Isolation of the Pyrrolizidine Alkaloid Europine N-oxide from Heliotropium Maris-mortui and h. Rotundifolium*. Phyrochemrsiry, 1978. **17**: p. 172
- 120. Lakshmanan, A.j. and S. shanmugasundaram, *Heliscabine, a Pyrrolizidine Ester Alkaloid from Heliotropium Scabrum*. Phytochemistry, 1995. **39**(2): p. 471-475.
- 121. Smith, L.W. and C.C.J. Culvenor, *Plant sources of hepatotoxic pyrrolizidine alkaloids*. J. Nat. Prod, 1981. **44**: p. 129-152.
- 122. Singh, B., et al., *Antineoplastic and antiviral screening of pyrrolizidine alkaloids from Heliotropium subulatum.* Pharm. Biol, 2002. **40**: p. 581-586.
- 123. Trigo, J.R., et al., *pyrrolizidine alkaloids in the acrtiid moth hyalurga syma*. J. Chem. Ecol, 1993. **4**: p. 669-679.
- 124. Roeder, E. and H. Wiedenfeld, *Pyrrolizidine alkaloids in plants used in the traditional medicine of Madagascar and the Mascarene islands.* Pharmazie, 2011. **66**: p. 637-647
- 125. Roeder, E., *Medicinal plants in China containing pyrrolizidine alkaloids*. Pharmazie, 2000. **55**(10): p. 711-26.
- 126. Wledenfeld, H., et al., *Pvrrolizidine Alkaloid containina Plants used in Monagolian Traditional Medicine: Lappula mvosotis Moench.* Scientia Pharmaceutica, 2005. **73**: p. 139-145.

- 127. Wiedenfeld, H., et al., *Pyrrolizidine Alkaloids from Lithospermum canescens Lehm.* Naturforsch, 2003. **58**: p. 173-176
- 128. FU, P.P., et al., *Pyrrolizidine Alkaloids Tumorigenic Components in Chinese Herbal Medicines and Dietary Supplements*. Journal of Food and Drug Analysis, 2002. **10**(4): p. 198-211.
- 129. RÖDER, E., et al., *Pyrrolizidine alkaloids of three taxa of Onosma (Boraginaceae-Lithospenneae)*. Phyton, 1993. **33**: p. 41-49.
- 130. El-Shazly, A., A. Abdel-Ghani, and M. Wink, *Pyrrolizidine alkaloids from Onosma arenaria (Boraginaceae)*. Biochemical Systematics and Ecology, 2003. **31**(5): p. 477-485.
- 131. Roeder, E., et al., *Pyrrolizidine alkaloids of three taxa of Onosma (Boraginaceae-Lithospermeae)*. Phyton, 1993. **33**: p. 41-49.
- 132. Roeder, E., et al., *Determination of open chain pyrrolizidine alkaloids by capillary gas chromatography.* Planta Med, 1990. **56**: p. 522.
- 133. Kretsi, O., et al., *Pyrrolizidine alkaloids from Onosma leptantha*. Helv.Chim. Acta, 2003. **86**: p. 3136-3140.
- 134. Mroczek, T., et al., On-line structure characterization of pyrrolizidine alkaloids in Onosma stellulatum and Emilia coccinea by liquid chromatography-ion-trap mass spectrometry. J. Chromatogr. A, 2004. **1056**: p. 91-97.
- 135. Liu, F., et al., Determination of pyrrolizidine alkaloids in comfrey by liquid chromatography–electrospray ionization mass spectrometry. Talanta, 2009. **80**(2): p. 916-923.
- 136. Semra Kurucu, M.K., M.I. Choudary, and G. Topcu, *Pyrrolizidine Alkaloids from Symphytum sylvaticum Boiss. subsp. sepulcrale. (Boiss. & Bal.) Greuter & Burdet var. sepulcrale and Symphytum aintabicum Hub. Mor. & Wickens.* Turk J Chem, 2002. **26** p. 195-199.
- 137. *Boraginaceae*, in *Flora of Libya*, S.M.H. Jafri and A. El-Gadi, Editors. 1979, Al Faateh University.
- 138. Ghani, A.E.-s.A.A., *Phytochemical Study Of Some Alkaloid Bering Plants*, in *Pharmacognacy*. 1990, Zagazig University: Zagazig.
- 139. Van Dam, N., et al., *The "Raison D'être" of pyrrolizidine alkaloids inCynoglossum officinale: Deterrent effects against generalist herbivores.* Journal of Chemical Ecology, 1995. **21**(5): p. 507-523.
- 140. Gardner, D.R., et al., *Pyrrolizidine alkaloids in Senecio madagascariensis from Australia and Hawaii and assessment of possible livestock poisoning*. Biochemical Systematics and Ecology, 2006. **34**(10): p. 736-744.
- 141. Mehrabani, M., et al., *Toxic Pyrrolizidine Alkaloids of Echium Amoenum Fisch. and Mey.* Daru Journal of Pharmaceutical Sciences, 2006. **14**(3): p. 122-127.
- 142. Christov, V., et al., *Alkaloids from the roots of Senecio macedonicus Griseb.* Z Naturforsch C, 2002. **57**(9-10): p. 780-4.
- 143. *The Challenges of Changing Retention Times in GC–MS*. 2007; Available from: <a href="http://www.chromatographyonline.com/">http://www.chromatographyonline.com/</a>.

- 144. Radecki, A. and J. Grzybowski, *Linear relationship between retention indices and chemical structure of phenols*. Journal of Chromatography A, 1978. **152**(1): p. 211-213.
- 145. Lee, J. and D.R. Taylor, *Relationships between temperature programmed and isothermal Kovats retention indices in gas-liquid chromatography*. Chromatographia, 1982. **16**(1): p. 286-289.
- 146. Kováts, E., Gas-chromatographische Charakterisierung organischer Verbindungen. Teil 1: Retentionsindices aliphatischer Halogenide, Alkohole, Aldehyde und Ketone. Helvetica Chimica Acta, 1958. **41**(7): p. 1915-1932.
- 147. El-Shazly, A.M., *Phytochemical Study of Some Alkaloid-Bering Plants*, in *Pharmacognocy*. 1995, university of Zagazig: Zagazig.
- 148. Wink, M. and L. Witte, *Storage of quinolizidine alkaloids in macrosiphum albifrons and aphis genistae homoptera aphididae*. Entomologia Generalis, 1991. **15**(4): p. 237-254.
- 149. Greinwald, R., et al., *Das Alkaloidmuster der Pfropfchimäre Laburnocytisus adamii* (*Fabaceae*). Biochemie und Physiologie der Pflanzen, 1991. **187**(5): p. 385-391.
- 150. Douglas Kinghorn, A., M.F. Balandrin, and L.-J. Lin, *Alkaloid distribution in some species of the papilionaceous tribes sophoreae, dalbergieae, loteae, brongniartieae and bossiaeeae.* Phytochemistry, 1982. **21**(9): p. 2269-2275.
- 151. El-Shazly, A., et al., *Pyrrolizidine Alkaloids from Echium setosum and Echium vulgare*. Journal of Natural Products, 1996. **59**(3): p. 310-313.
- 152. El-Shazly, A., et al., *Pyrrolizidine alkaloids of Cynoglossum officinale and Cynoglossum amabile (family boraginaceae)*. Biochemical Systematics and Ecology, 1996. **24**(5): p. 415-421.
- 153. El-Shazly, A., A. Abdel-Ghani, and M. Wink, *Pyrrolizidine Alkaloids from Onosma arenaria Waldst. and Kit.*(*Boraginaceae*). Biochemical Systematics and Ecology, 2003. **31**: p. 477-485.
- 154. Kelley, R.B. and J.N. Seiber, *Pyrrolizidine alkaloid chemosystematics in Amsinckia*. Phytochemistry, 1992. **31**(7): p. 2369-2387.
- 155. Pedersen, E. and E. Larsen, *Mass spectrometry of some pyrrolizidine alkaloids*. Organic Mass Spectrometry, 1970. **4**(S1): p. 249-256.
- 156. Dannhardt, G., *Pyrrolizidin-Alkaloide: Chemistry and Toxicology of Pyrrolizidine Alkaloids. Von A. R. Mattocks. Academic Press, London Orlando San Diego New York Austin Montreal Sydney Tokyo Toronto 1986. 393 S., Tab. £ 55.00. ISBN 0-12-480570-1.* Nachrichten aus Chemie, Technik und Laboratorium, 1987. **35**(1): p. 41-41.
- 157. Stelljes, M.E., et al., *GC-MS Determination of Pyrrolizidine Alkaloids in Four Senecio Species.* Journal of Natural Products, 1991. **54**(3): p. 759-773.
- 158. El-Shazly, A., et al., *Pyrrolizidine alkaloids from Echium rauwolfii and Echium horridum (Boraginaceae)*. Z Naturforsch C, 1999. **54**(5-6): p. 295-300.
- 159. Nowacki, E.K. and G.R. Waller, *Metabolism of l-sparteine, dl-lupanine and l-thermopsine in species of Leguminosae*. Phytochemistry, 1975. **14**(1): p. 161-164.

- 160. Roeder, E. and T. Bourauel, *Pyrrolizidine alkaloids from Neatostema apulum*. Phytochemistry, 1992. **31**(10): p. 3613-3615.
- 161. Witte, L., et al., *Comparative analysis of pyrrolizidine alkaloids from natural sources by gas chromatography-mass spectrometry*. Phytochemistry, 1992. **32**(1): p. 187-196.
- 162. Rashkes, Y.V., U.A. Abdullaev, and S.Y. Yunusov, *Mass spectra of pyrrolizidine alkaloids*. Chemistry of Natural Compounds, 1978. **14**(2): p. 121-135.
- 163. John, A.E. and W.S. Leslie, *Transfer of Pyrrolizidine Alkaloids into Eggs: Food Safety Implications*, in *Natural and Selected Synthetic Toxins*. 1999, American Chemical Society. p. 118-128.

#### الملخص

هذه الدراسة تتضمن عرض مختصر لقلويدات البيروليزيدين التي تم فصلها سابقا من أصناف متعددة لنباتات من العائلة الشفوية باستخدام تقنيات مختلفة

الهدف الأساسي لهذا البحث هو فصل وتحديد تركيب قلويدات البير وليزيدين من مستخلص الميثانولي لنبات Echium plantagenium (يعرف محليا باسم المصيص) الذي يعود للعائلة الشفوية والذي ينمو بالقرب من مدينة المرج حيث تم فصل أثنى عشر مركبا:

#### Ech-7

7-Angeloyl-9-(2,3-dihydroxybutyryl) retronecine

Ech-8 and Ech-9	Ech-10	
Assumed to be isomer of Ech-7	HO HO HO Uplandicine	
	Opiandienie	
Ech-11	Ech-12	
HO HO HO Echimidine	HO OH OH OHO Echiumine	

تم التعرف على المركبات المفصولة باستخدام تقنية Retention Indices (RI) و كذلك حساب قيم Spectrometry حيث تم مقارنة النتائج المتحصل عليها مع نتائج اخرى لمركبات مفصولة سابقا كقيم معايرية.